

Network connectivity is shown to change in C57BL/6 mice during a continuing immune response subsequent to tetanus toxoid hyperimmunization

Irena P Zivkovic¹, Marijana M Stojanovic¹, Vladimir Z Petrusic¹, Aleksandra B Inic-Kanada¹, Mileva V Micic² & Ljiljana A Dimitrijevic¹

¹ Institute of Virology, Vaccines and Sera - Torlak, Department of Research and Development, Vojvode Stepe 458, 11152 Belgrade, Serbia;

² Institute for Medical Research, Dr Subotica 4, PO Box 102, 11129 Belgrade, Serbia.

ABSTRACT

We have already demonstrated (Stojanovic et al., 2009) a connection between tetanus toxoid (TTd) hyperimmunization and the induction of anti-phospholipid syndrome (APS) in BALB/c mice. Here we show that C57BL/6 mice subjected to an identical procedure do not exhibit any like pathology attributable to anti-phospholipid antibodies; we explain that this absence results from idiotypic connectivity. Six groups of C57BL/6 mice were hyperimmunized with TTd in aluminum hydroxide or glycerol, with or without pretreatments. Pretreated mice had been injected with polyclonal or nonspecific immune stimulators, such as complete Freund's adjuvant (CFA) or glycerol. The epitope specificity of induced antibodies was tested by indirect ELISA using a tetanus toxoid immunogen and these autoantigens: phospholipids, gangliosides, laminin. Idiotypic connectivity was tested by competitive ELISA and gauged from the degree to which the interaction of idiotypic/anti-idiotypic complementary antibodies was inhibited in the presence of immunized sera antibodies. Higher idiotypic connectivity was noted amongst pretreated mice. There was a positive correlation between higher connectivity and autoantibody levels that acted to favor the participation of natural autoantibodies in the inhibitory process. We conclude that idiotypic connectivity plays a protective role in immunization-induced autoimmunity.

Key terms: Autoantibodies, C57BL/6 mice, hyperimmunization, network

INTRODUCTION

Of the mechanisms that may contribute to autoantibody (AAb) generation, those most often cited are molecular mimicry (Croxford et al., 2005, Samarkos & Vaiopoulos, 2005) and polyclonal cell activation (Wickham & Carr, 2004, Ramos et al., 2005). Antibodies (Abs) generated during immunization, whether immunogen- or self-specific, may have either public or private idiotypes (Ids) (Reilly & Root, 1986, Hebert et al., 1990). These two Ids generate anti-idiotypic (anti-Id) Abs, respectively regarded as either autoantigens or AAbs; a network of interacting Ids and anti-Id Abs is thereby formed in consequence (Jerne, 1974, 2004 b, Vani et al., 2009). It is not suggested that the network potentially created by Id/anti-Id interactions following immunization is regulatory in nature; this is at variance with the network of Id/anti-Id interactions between natural autoantibodies (NAAbs) with public Id (Vani et al., 2008, Varela & Coutinho, 1991, Coutinho, 1995). Since anti-Ids may be thought of as virtual AAbs, it is valid in this context to invoke the mechanisms of AAb induction when analyzing changes (be they qualitative or quantitative) in connectivity between Ids and anti-Ids induced during immunization, these mechanisms being selective polyclonal B cell expansion (Sutmuller et al., 1999) and/or an antigen-driven process (Shoenfeld, 1994, Atassi & Casali, 2008) followed by mutations and strong Abs selection. Altered connectivity between Ids and anti-Ids may perhaps be the *primum movens* of autoimmune diseases; the loss of Id connectivity in the course of autoimmune diseases has been variously reported (Rossi et al., 1990, Very et al., 1993, Dwyer et al., 1986,

Pacheco-Garcia et al., 1999, Poletaev, 2002). The demonstratedly beneficial therapeutic effect of intravenous immunoglobulin (IVIG) administered to patients suffering from autoimmune disease, though its mode of operation is yet to be fully elucidated, lends credence to its importance in the restoration of Id connectivity (Dalakas, 2004, Kazatchkine & Kaveri, 2001, Bayary et al., 2006).

In this paper we analyze changes in natural Id expression and AAb connectivity in the sera of C57BL/6 mice hyperimmunized with tetanus toxoid (TTd) in conformity with six immunization protocols. TTd was the immunogen of choice because it is a partial structural homolog of beta2-glycoprotein I (β 2GPI), a major autoantigen in antiphospholipid syndrome (APS) (Blank et al., 2002); furthermore, we have already demonstrated a link between TTd hyperimmunization and APS induction in BALB/c mice, when the severity of the pathology (fetal resorptions) was shown to be TTd administration context dependent (Stojanovic et al., 2009). The pathological response found in TTd-hyperimmunized BALB/c mice is in stark contrast to the case of C57BL/6 mice where this type of reproductive pathology has not been observed. It is hypothesized that the absence (whatever the context of TTd administration) of this characteristic reproductive pathology subsequent to immunization in the C57BL/6 strain may be explained by a synchronized elevation of highly connected AAbs, which curb the effects of any pathological antibodies that may be present.

These were our hyperimmunization protocols: a hyperdose of TTd in either aluminum hydroxide or glycerol adjuvants was injected into non-, glycerol or CFA-pretreated C57BL/6 mice; we thus generated six groups of dissimilarly

immunized animals from the six possible permutations of adjuvant and pretreatment.

Antibody profiles were studied over a period of 22 weeks with nine blood sera samples being taken at two-week intervals: preliminary samples of untreated sera were taken immediately before pretreatments, one week before the beginning of the course of immunizations; eight further samples of treated blood sera were taken every two weeks thereafter; a tenth and final sample was taken one week after the administration of the TTd booster dose. We were monitoring for changes in: Id/anti-Id connectivity; reactivity towards immunogen (TTd) and autoantigens (β 2GPI, phospholipids, gangliosides and laminin); isotype composition.

The expression of natural Ids on TTd-immunized mouse Abs during the course of immunization was tracked via the inhibition of interaction in a complementary idiotype pair: idiotype IgM DJ and its specific anti-idiotype murine monoclonal antibody (MoAb) Y7.

The Y7 idiotype was first described occurring on human IgM molecules (IgM DJ) and then classified as natural. Natural, firstly, from its expression, detected on 5 % to 8 % of IgM from cord blood sera (Radulovic et al., 1997) and from the sera of patients suffering chronic lymphocytic leukemia (Ivanovic et al., 1990). Natural, secondly, from its IgM DJ properties, binding to ssDNA, lactic acid bacteria (LAB) and phosphorylcholine (PC) passively adsorbed to bovine serum albumin (BSA) (Dimitrijevic et al., 1999). The Y7 idiotype was assigned to the subgroup V_H3 (Dimitrijevic et al., 2004 a).

The anti-idiotype property of MoAb Y7 was characterized by its ability to inhibit IgM DJ immunogen binding to PC adsorbed to BSA (Dimitrijevic et al., 1992). This ability, indicative of internal image properties, leads to its typing as an analog of Ab2-beta. It may be regarded as an internal image of the natural idiotype. That the Y7 idiotype/MoAb Y7 pair has a regulatory function in a murine immune system was shown from its property of shaping the immune response against LAB in BALB/c mice (Dimitrijevic et al., 2004 b).

Our results show that the pretreatments shaped the profiles of analyzed AAbs, tending to the hypothesis that polyclonal-induced antibody Id interactions participated in both the pathology and in the control of the immune response. The absence in the C57BL/6 strain of the characteristic reproductive pathology (fetal resorption) found earlier in BALB/c mice consequent to immunization (Stojanovic et al., 2009) may be explained by a synchronized elevation of these highly-connected AAbs neutralizing any threat posed by potentially pathological Abs.

METHODS

Immunization and bleeding schedules

10-week-old C57BL/6 female mice were used in the experiments. Pretreatments (a single dose of CFA or 2.5 M glycerol (glyc) injected subcutaneously) were performed one week before the first administration of TTd. Non-pretreated, CFA-pretreated or glyc-pretreated mice were immunized with high doses of TTd (Institute of Virology, Vaccines and Sera - Torlak, Belgrade) mixed with either glyc (TTd/glyc) or 2 % aluminum hydroxide (Al; TTd/Al) as adjuvants. Doses

of 200 μ l per mouse were administered subcutaneously three times at two-week intervals, 500 μ g/ml TTd per 200 μ l dose. 16 weeks after the third dose, one additional booster dose of TTd was administered with an appropriate adjuvant. Thus we generated six groups of dissimilarly immunized animals (10 mice per group) from the six possible permutations of adjuvant and pretreatment.

The TTd used for immunization had passed the tests of specific and reverse toxicity to meet the requirements of the *European Pharmacopoeia*.

Samples of normal blood sera were collected simultaneously from all groups by bleeding from the retro-orbital plexus prior to any intervention, i.e. immediately before pretreatments. Subsequent samples were similarly taken at two-week intervals for 16 weeks followed by one final sample taken one week after the administration of the TTd booster dose. Each group therefore underwent 10 separate bleedings inclusive of the initial taking of normal sera. The general methodology of our immunization and bleeding schedules has been previously described (Stojanovic et al., 2009; *vide* REFERENCES 36. *infra*).

The collected sera were complement depleted, aliquoted and stored at -20 °C until used for analyses. The sample sera pools as assayed contained identical integrated volumes of sera, taken at identical time points (relative to the start of treatment) from identically immunized mice.

All animal experimentation was conducted in accordance with our country-specific *Guiding Principles for the Care and Use of Laboratory Animals*; these guidelines conform with the provisions of the Declaration of Helsinki (as revised and clarified); our experimentation received the approval of the Animal Institutional Care and Use Committee at the Institute of Immunology and Virology - Torlak.

Indirect ELISA for the detection of specific sera IgG populations

Detection of sera Abs specific for TTd, β 2GPI, phospholipids or laminin was performed by indirect enzyme-linked immunosorbent assay (ELISA). Optimal sera dilutions were defined according to the results of preliminary experiments where randomly selected sera were analyzed at various dilutions. The sera Abs' reactivity towards specific antigens (Ags) was simultaneously analyzed in all discrete samples.

Ag-specific sera IgG binding was detected by biotin-labeled anti-mouse IgG (*Sigma-Aldrich*, St. Louis, Missouri, USA) or by using commercially available biotin-labeled Abs specific for murine IgG subclasses: anti-mouse IgG1, anti-mouse IgG2b, and anti-mouse IgG3 (ICN Biomedicals Inc., Aurora, Ohio, USA); anti-mouse IgG2c (Jackson ImmunoResearch Laboratories, West Grove, Pennsylvania, USA). The commercially available Abs were used in accordance with the manufacturers' instructions and incubated at room temperature (1 h). In all cases the ExtrAvidinTM-peroxidase/o-phenylenediamine (OPD) system (*Sigma-Aldrich Chemie GmbH*, Steinheim, Germany) was used to visualize Ag-Ab interactions. Absorbance was monitored at 492 nm and 620 nm ($A_{492/620}$). The cut-off value for each system was defined according to the $A_{492/620}$ reading in negative control wells containing samples 1 % w/v BSA in phosphate-buffered saline (PBS) + 3 \times SD. Sera samples were considered positive when the $A_{492/620}$ value exceeded the cut-off value.

Detection of IgG specific for TTd and β 2GPI

The TTd used in ELISA had been produced at the Institute of Virology, Vaccines and Sera - Torlak, Belgrade. β 2GPI was isolated from human plasma by a combination of precipitation techniques and ion-exchange chromatography (Stojanovic et al., 2009). Commercially available mouse anti-human β 2GPI IgG1, clone 5F7, (ICN Biomedicals Inc., Aurora, Ohio, USA) was used for the identification of protein isolates.

Nunc MaxiSorp™ ELISA plates (Nunc, Roskilde, Denmark) were covered (50 μ l/well) with TTd (1 μ g/ml TTd in PBS) or β 2GPI (10 μ g/ml β 2GPI in PBS) by overnight adsorption (4 °C). 1 % w/v BSA in PBS was used for blocking at room temperature (2 h). Blocking and all subsequent steps in the ELISA were followed by washing with 0.05 % (v/v) Tween 20 in PBS (four times, 200 μ l/well). Appropriately diluted (1:800 in 1 % w/v BSA in PBS) sera samples were incubated at room temperature (1h, 50 μ l/well).

Detection of IgG specific for mouse laminin

Mouse laminin (Sigma-Aldrich, St. Louis, Missouri, USA) (10 μ g/ml, 50 μ l/well) was adsorbed onto Nunc MaxiSorp™ microtiter plates (4 h, 37 °C). The plates were then washed (three times, 200 μ l/well PBS) and blocked with 1 % w/v BSA in PBS. Diluted (1:400 in 1 % w/v BSA in PBS, 0.05 % v/v Tween 20 in PBS) sera samples were added to the plates after washing four times (200 μ l/well, 0.05 % v/v Tween 20 in PBS) and then incubated overnight at 4 °C.

Detection of IgG specific for phospholipids or gangliosides

For these ELISAs commercially available phospholipids and gangliosides (Sigma-Aldrich Chemie GmbH, Steinheim) were used as antigens. The reactivities of sera IgG to the following phospholipids were analyzed: phosphatidylcholine (PtC), phosphatidylserine (PS), phosphatidylethanolamine (PE), phosphatidylinositol (PI), phosphatidylglycerol (PG) and cardiolipin (CL). Disialoganglioside G_{D1b} and trisialoganglioside G_{T1b} were used as antigens for the evaluation of specific anti-ganglioside sera IgG levels.

Phospholipids (10 μ g/ml in absolute ethanol, 50 μ l/well) or gangliosides (10 μ g/ml in methanol, 50 μ l/well) were immobilized onto Nunc PolySorp™ ELISA plates by evaporation at room temperature. After washing unbound Ags (three times, 200 μ l/well PBS) and blocking of free sites (200 μ l 1 % w/v BSA in PBS; 1 h, 37 °C), the diluted (1:400 in 1 % w/v BSA in PBS, 50 ml/well) mice sera were added. To detect anti-phospholipid IgG the sera samples were incubated at room temperature (1 h); to detect anti-ganglioside IgG the sera samples were incubated overnight (4 °C). All subsequent ELISA steps were followed by washing with 0.05 % v/v Tween 20 in PBS (four times, 200 μ l/well).

Competitive ELISA for the detection of Y7 natural Id concentration changes in the sera of immunized mice

Two MoAbs were used to measure Y7 expression on sera Abs: biotin-labeled F(ab)₂ of the anti-idiotypic murine MoAb Y7 (F(ab)₂ Y7-B) and Y7⁺ human MoAb IgM DJ. The Y7 concentration was estimated from the inhibition of F(ab)₂ Y7-B binding to IgM adsorbed onto plastic. IgM DJ was bound

to a Nunc Maxisorp™ microtiter plate (50 μ l/well) at a concentration of 500 ng/ml. F(ab)₂ Y7-B (200 ng/ml) was mixed with mouse sera diluted in 1 % w/v BSA in PBS to a final dilution of 1:100, preincubated (1 h, 25 °C) in a water bath and further incubated with coated IgM DJ. ExtrAvidin™-peroxidase/OPD was used as a detector. Percentage inhibition (PI) was calculated after the determination of free F(ab)₂ Y7-B using the standard curve of binding to IgM DJ.

RESULTS

TTd-specific (Ab1) and anti-ganglioside (Ab2) IgG antibodies

Time-dependent fluctuations in anti-TTd and anti-ganglioside sera IgG levels in TTd/Al and TTd/glyc-immunized C57BL/6 mice are shown (Fig. 1 and Fig. 2). No matter which protocol was followed, it was found that TTd hyperimmunizations induced an intensive production of anti-TTd specific IgG. A challenge test with tetanus toxin confirmed that a protective immune response had developed in all groups (data not shown). In the context of Id networking, these TTd-specific Abs may be regarded as type Ab1. In all groups, a dramatic rise in sera anti-TTd IgG levels was observed during the main immunization phase, peaking at week six (two weeks after the administration of the third dose) and declining thereafter. Our results show that any difference in the binding characteristics and dynamics of these immunogen-specific sera IgG Abs was primarily a function of the adjuvant. The highest levels of immunogen-specific IgG were produced following TTd immunization in the presence of adjuvant Al; levels were lower in an identically pretreated group immunized in the presence of adjuvant glyc. In the immediate post-immunization phase (prior to booster administration), anti-TTd IgG levels were found to decline with varying degrees of rapidity in mice immunized with TTd/glyc; in TTd/Al immunized mice, sera levels of TTd-specific IgG remained stable.

Following the booster TTd dose, however, a marked rise in the production of anti-TTd IgG was observed only in those immunizations administered in the presence of adjuvant glyc; anti-TTd IgG levels following immunizations given in the presence of adjuvant Al had already reached their plateaux and remained stable at those levels even after the final TTd booster dose.

In each group the increase in anti-TTd IgG sera levels was followed by an increase in anti-ganglioside IgG production. G_{D1b} and G_{T1b} are receptors for the tetanus toxin; from this it follows that the Ag binding site of anti-ganglioside IgG must be an internal image of the anti-TTd Ab reactive to the ganglioside binding site found on the tetanus Ag. In the context of idiotypic networks, anti-ganglioside Abs may be regarded as type Ab2. A comparison of time-dependent fluctuations in the levels of anti-TTd IgG and anti-ganglioside IgG confirmed that in all groups the appreciable rise in anti-TTd IgG production preceded the appreciable rise in anti-ganglioside IgG production. However, the most pronounced rises in anti-ganglioside Ab levels were seen during the post-immunization period (regardless of the immunization protocol followed), differing in this respect from the observed patterns of increase in TTd-specific IgG levels.

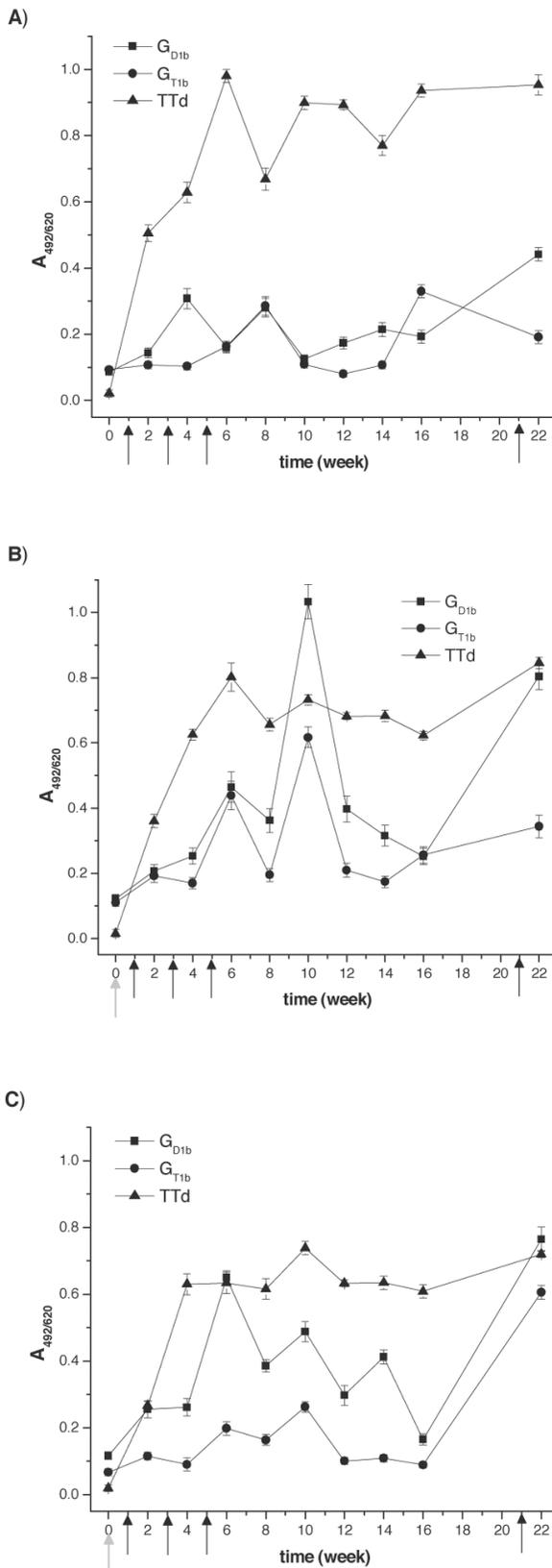


Figure 1: Time-dependant fluctuations in anti-TTd, anti- G_{D1b} and anti- G_{T1b} IgG levels within sera of non-pretreated (A), CFA-pretreated (B) or glyc-pretreated (C) C57BL/6 mice and hyperimmunized by TTd/Al. Light grey arrows indicate pretreatments and black arrows indicate TTd injections. Each sample (defined sera pool) was assayed in triplicate and calculated mean $A_{492/620} \pm SE$ are presented.

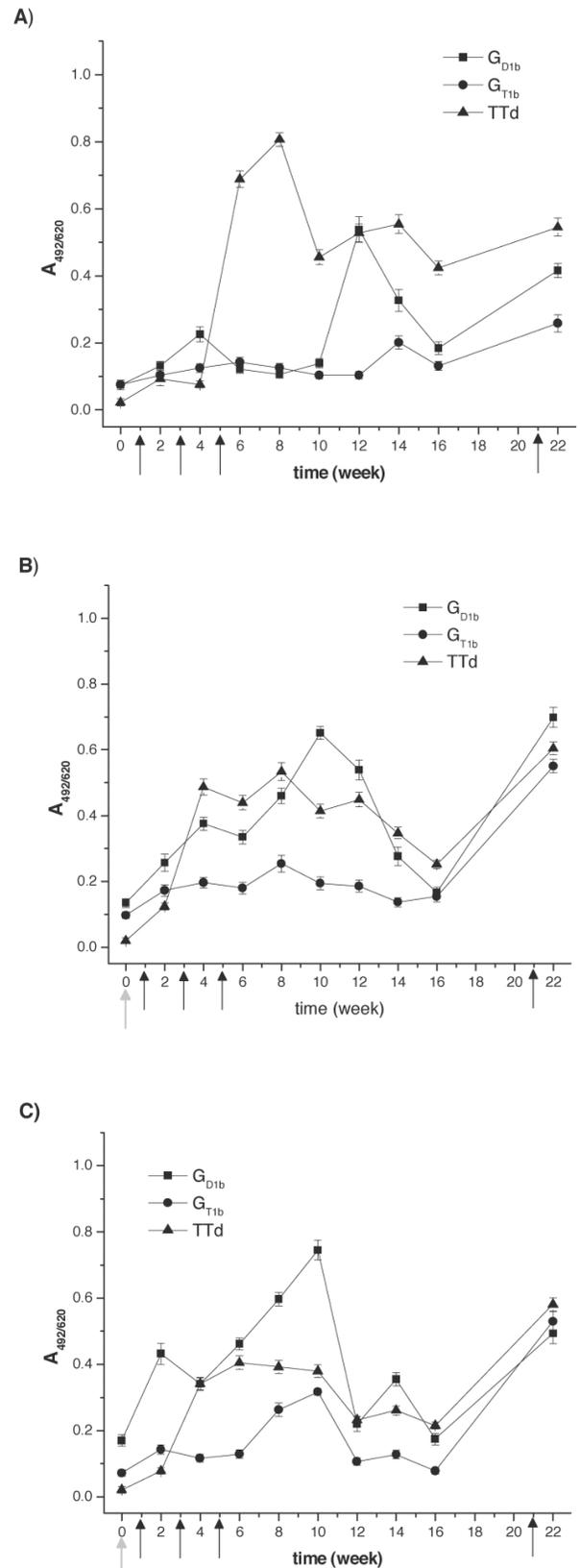


Figure 2: Time-dependant fluctuations in anti-TTd, anti- G_{D1b} and anti- G_{T1b} IgG levels within in sera of non-pretreated (A), CFA-pretreated (B) or glyc-pretreated (C) C57BL/6 mice and then hyperimmunized by TTd/glyc. Light grey arrows indicate pretreatments and black arrows indicate TTd/glyc injections. Each sample (defined sera pool) was assayed in triplicate and calculated mean $A_{492/620} \pm SE$ are presented.

The levels of anti-ganglioside Abs were for the most part found to be higher among pretreated (be it with CFA or glyc) TTD-immunized C57BL/6 mice than among their non-pretreated TTD-immunized counterparts. Likewise, anti-ganglioside IgG production in response to the booster TTD dose was more intensive among pretreated mice.

Analyses of specific IgG subclass contributions to the entirety of the anti-ganglioside IgG pools showed that, for all experimental groups, a significant moiety of these contributing Abs belonged to subclasses IgG1 and IgG2c. The immunizations had no significant influence on either the dynamics or the contribution levels of any one specific anti-ganglioside IgG subclass; the analyses (Fig. 3) of the subclasses within anti- G_{D1b} and anti- G_{T1b} sera IgG pools (here shown as found in CFA-pretreated and TTD/glyc immunized mice only) are representative of all other mouse groups.

Anti-phospholipid, anti- β 2GPI and anti-laminin IgG antibodies

The reactivity of TTD-hyperimmunized sera IgG to the autoantigens normally recognized by NAAs (phospholipids, β 2GPI) was analyzed; sera reactivity to autoantigens bearing a partial structural similarity to the tetanus toxin/toxoid was analyzed by laminin or β 2GPI.

Fluctuations in the levels of anti- β 2GPI, anti-phospholipids and anti-laminin IgG AAbs following specific immunizations were plotted against time (Fig. 4A and 4B; Fig. 5A and 5B; Fig. 6A and 6B).

Time-dependent changes in the reactivity of sera IgG to individual phospholipids were near identical, whatever the phospholipid type. In the context of NAAs, anti-PtC is protective against infection and anti-PS IgG is active in the removal of apoptotic cells; because of these functions, they are the best explored of the phospholipids and so we have chosen PtC and PS to stand for phospholipids as a whole.

We have shown that TTD-hyperimmunization initiates production of significantly elevated levels of both anti- β 2GPI IgG and phospholipid-specific IgG. Regardless of group,

maximal sera levels of these AAbs were detected during the post-immunization period; however, the dynamics of production and the levels attained within individual groups were a function of the pretreatments administered. In pretreated mice, irrespective of adjuvant, anti- β 2GPI and anti-phospholipid IgG reached maximal sera levels between the eight and tenth weeks following the course's commencement (Fig. 4A and 4B; Fig. 5A and 5B); in non-pretreated mice these concentrations rose gradually over a period of eight weeks during the post-immunization follow-up (Fig. 6A and 6B). The maximal attained sera concentrations of anti- β 2GPI and anti-phospholipid IgG (evaluated from $A_{492/620}$) were not adjuvant-dependent, ranking (highest to lowest): CFA-pretreated > glyc-pretreated > non-pretreated. The shapes of curves depicting time-dependent fluctuations in the level of defined IgG AAbs (anti- β 2GPI and anti-phospholipid) were broadly similar for all the experimental groups (A and B on Fig. 4, Fig. 5 and Fig. 6).

No significant time-dependent fluctuations in anti-laminin IgG pools were seen in any of the experimental groups (A and B on Fig. 4, Fig. 5 and Fig. 6).

The binding of Y7 F(ab)₂ fragments to IgM DJ in the presence of TTD-hyperimmunized mice sera

The probable influence of the previously described immunization procedures on Id/anti-Id interactions and on network perturbation was gauged from the extent to which the binding of anti-Id MoAb Y7 F(ab)₂ to its corresponding Id (Y7⁺ IgM DJ) was inhibited in the sera of the TTD-immunized C57BL/6 mice.

Levels of NAAs vary, as one might expect given the broad physiological spectra across which they manifest (PI of normal sera pools was 32-50%); over any extended period of time these levels can be expected to oscillate by as much as 20 %, thus only fluctuations exceeding this 20 % bound were deemed significant. All immunization protocols induced

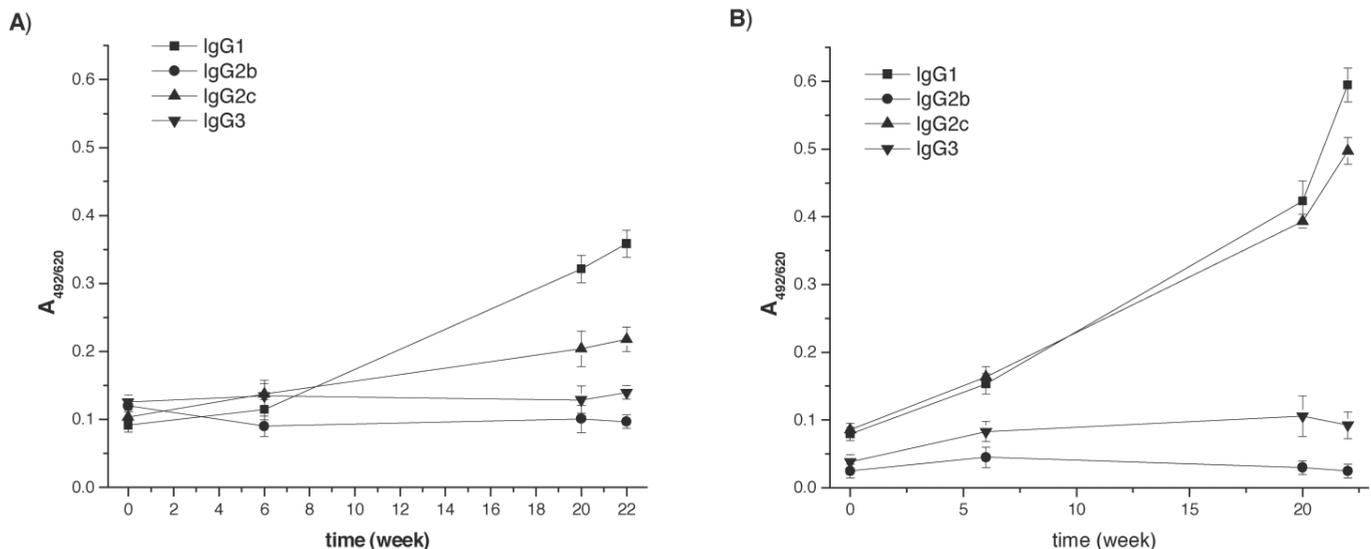


Figure 3: Influence of CFA-pretreatment and subsequent hyperimmunization by TTD/glyc on IgG subclasses levels within sera IgG pools specific for G_{D1b} (A) and G_{T1b} (B). Each sample (defined sera pool) was assayed in triplicate and calculated mean $A_{492/620} \pm$ SE are presented.

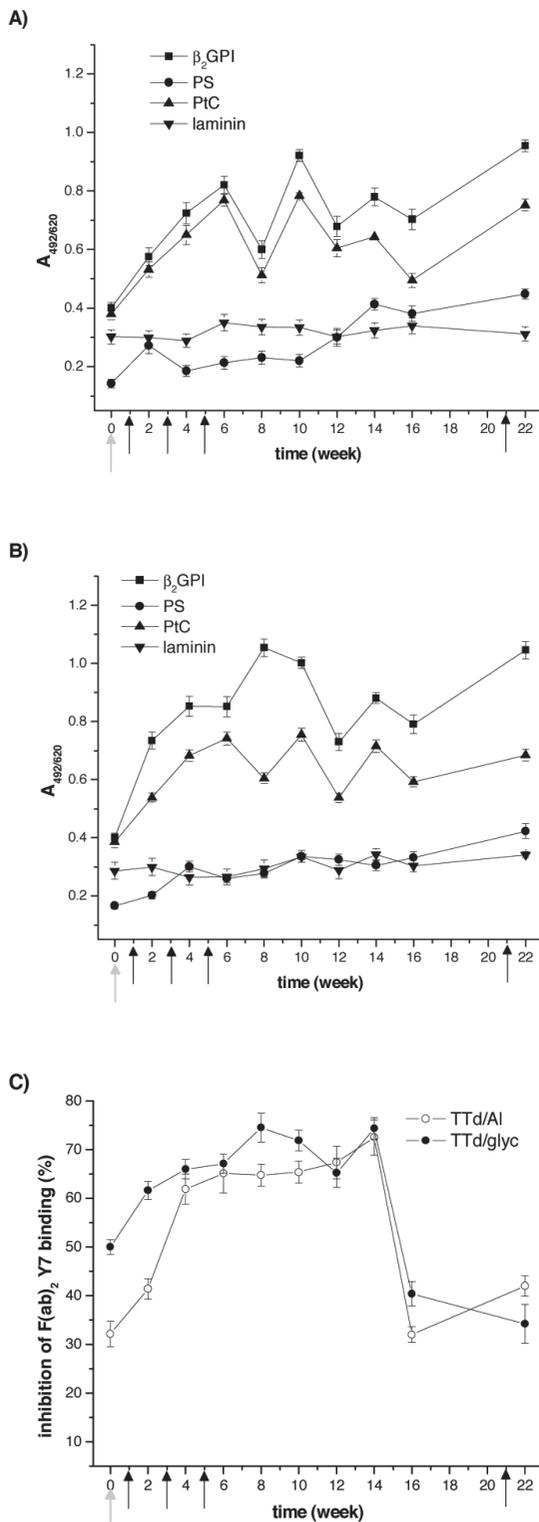


Figure 4: Time-dependent fluctuations in anti-B2GPI, anti-laminin, anti-PS and anti-PtC IgG levels within sera of glycopretreated C57BL/6 mice and hyperimmunized by TTd/Al (A) or by TTd/glyc (B). Each sample (defined sera pool) was assayed in triplicate and mean $A_{492/620} \pm SE$ are shown. Chart C plots inhibition of $F(ab)_2$ Y7 binding to IgM DJ (Y7+ IgM) as measured in sera pools collected at defined time-points from glycopretreated C57BL/6 mice immunized by TTd/Al or TTd/glyc. Each sample sera pool was assayed in triplicate and calculated mean PI $\pm SE$ are presented. Light grey arrows indicate pretreatment and black arrows indicate TTd injections.

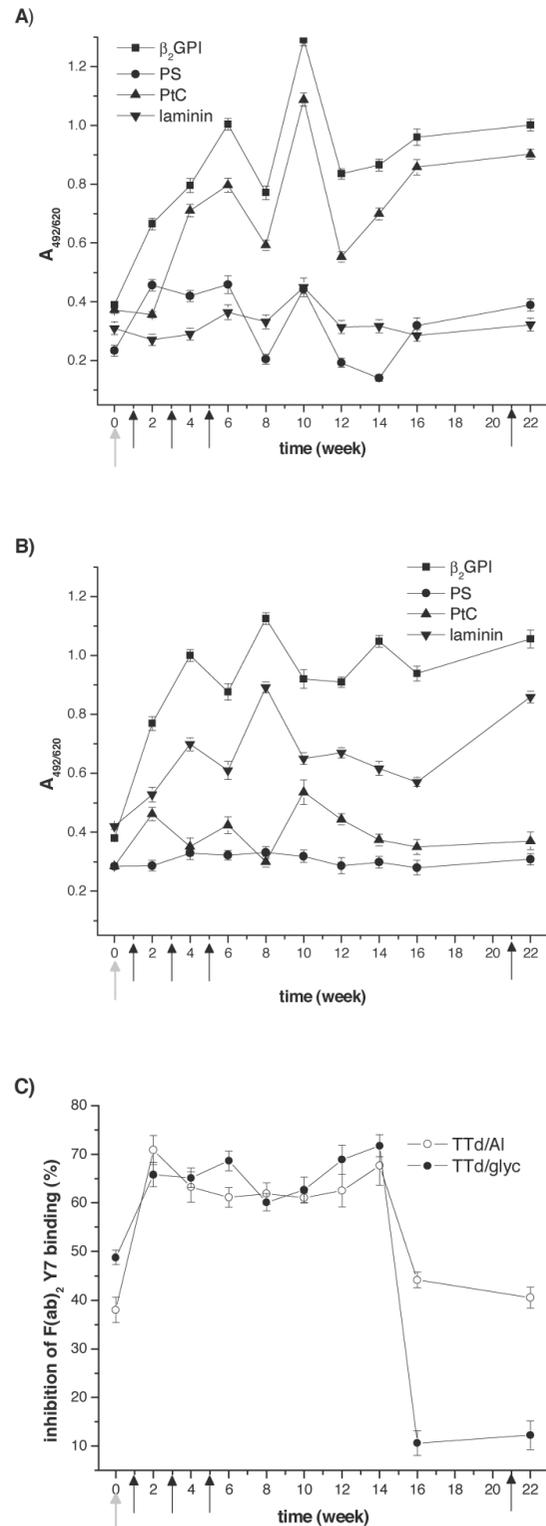


Figure 5: Time-dependent fluctuations in anti-B2GPI, anti-laminin, anti PS and anti-PtC IgG levels within sera of CFA-pretreated C57BL/6 mice and hyperimmunized by TTd/Al (A) or by TTd/glyc (B). Each sample (defined sera pool) was assayed in triplicate and mean $A_{492/620} \pm SE$ are shown. Chart C plots inhibition of $F(ab)_2$ Y7 binding to IgM DJ (Y7+ IgM) as measured in sera pools collected at defined time-points from CFA pretreated C57BL/6 mice immunized by TTd/Al or TTd/glyc. Each sample sera pool was assayed in triplicate and calculated mean PI $\pm SE$ are presented. Light grey arrows indicate pretreatment and black arrows indicate TTd injections.

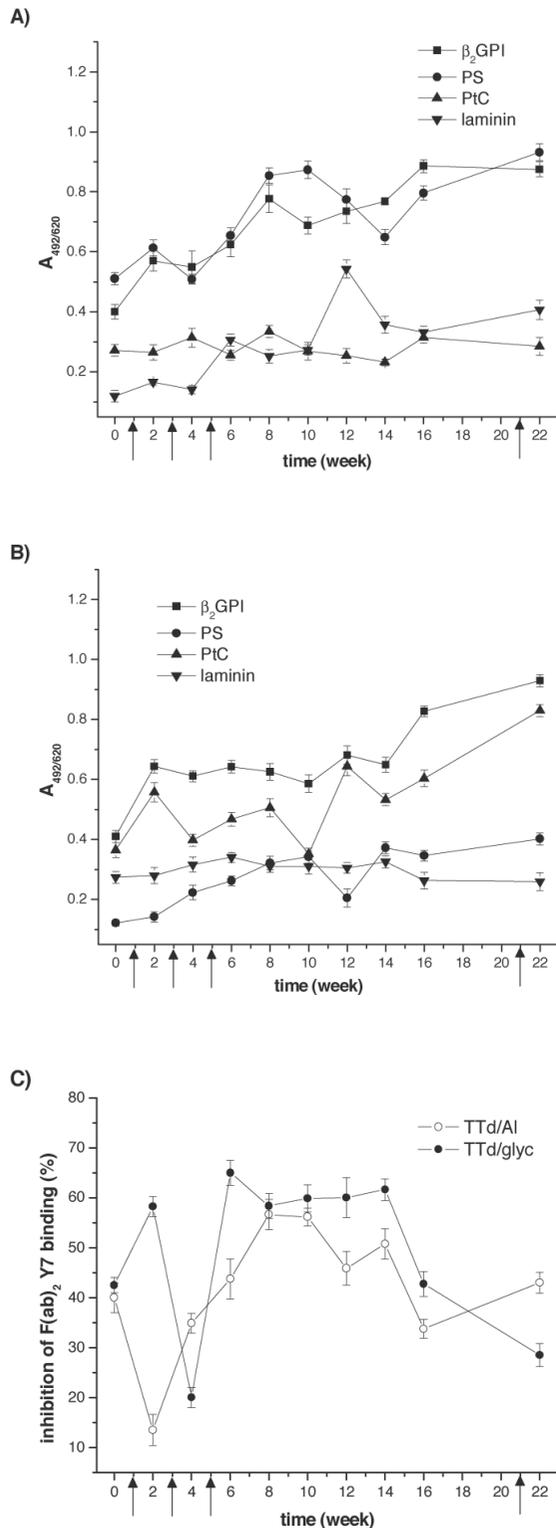


Figure 6: Time-dependent fluctuations in anti-B2GPI, anti-laminin, anti-PS and anti-PTC IgG levels within sera of non-pretreated C57BL/6 mice and hyperimmunized by TTd/Al (A) or by TTd/glyc (B). Each sample (defined sera pool) was assayed in triplicate and mean $A_{492/620} \pm$ SE are shown. Chart C plots inhibition of $F(ab)_2$ Y7 binding to IgM DJ ($Y7 +$ IgM) as measured in sera pools collected at defined time-points from non-pretreated C57BL/6 mice immunized by TTd/Al or TTd/glyc. Each sample sera pool was assayed in triplicate and calculated mean PI \pm SE are presented. Black arrows indicate TTd injections.

intensive fluctuations within NAAb pools conformant with the inhibition of MoAb Y7 $F(ab)_2$ - IgM DJ binding (Fig. 4C, Fig. 5C and Fig. 6C). Whichever immunization protocol was followed, sera collected during the post-immunization period were more potent inhibitors of MoAb Y7 $F(ab)_2$ - IgM DJ interactions than were any of the normal C57BL/6 sera collected prior to intervention. The potential of sera Abs to inhibit MoAb Y7 $F(ab)_2$ - IgM DJ interactions was found to be time-dependent; a close analysis of the changes in this inhibitory potential revealed that network perturbations were more intensive and occurred more rapidly amongst pretreated mice than amongst non-pretreated mice. The adjuvant used not being a criterion, sera found to have the greatest inhibitory effect on MoAb Y7 $F(ab)_2$ - IgM DJ interactions (and the degree of effect did have to be significantly in excess of that produced by normal sera) were collected: in the case of CFA-pretreated mice after the first TTd dose; from glyc-pretreated mice after the second dose; from non-pretreated mice only at the very end of the immunization protocol.

For each group, the sera's marked inhibitory potential persisted until the ninth week post-immunization; its inhibitory potential then fell abruptly and was not significantly restored by the final TTd-booster dose administration.

DISCUSSION

In this paper we show that, in C57BL/6 mice, TTd hyperimmunization (whichever of our chosen protocols governed the immunizations) induces a high level of immunogen specific IgG production. There was a concomitant rise in the reactivity of sera IgG towards various autoantigens (AAgs), accompanied by quantitative disturbances in preexisting Id/anti-Id interactions demonstrated by the inhibition of MoAb Y7 and $Y7 +$ IgM DJ binding.

Our results show that the TTd-specific and the autoreactive immune responses (which developed in tandem) are differently regulated: the immunogen-specific immune response was primarily shaped by the adjuvant used (whether Al or glyc); the dynamics and the intensities of the self-directed immune responses were linked to the pretreatments administered.

TTd-hyperimmunization in all groups induced a significant rise in reactivity towards the AAgs commonly recognized by NAAbs, as gangliosides, β_2 GPI and phospholipids. The reactivity of sera IgG towards mouse laminin (rarely recognized by mouse NAAbs) did not alter significantly within any experimental group, a result which, whilst implying that murine laminin tolerance is strictly regulated, is unsurprising given that anti-laminin IgGs are overtly pathogenic in the murine system. This invariance in reactivity accords with recent data showing that the robustness of any tolerance mechanism is AAg-specific and correlates inversely with the contributions of specific AAbs to the pool of NAAbs (Wang et al., 2003).

That autoimmunity is associated with the triggering of increases in AAb production and disturbances in Id networks is known; the trigger itself is not. Whether the initiating process is driven by antigens, by polyclonal B cell activation, or by both of these acting in concert, is as yet unelucidated but either or both are plausible candidates.

Our pretreatments (CFA, glyc) and adjuvants (Al, glyc) are all potent polyclonal stimulators. Aluminum may be known, first and foremost, as a B cell stimulator but it also influences Ag-presenting cell activity (Brewer, 2006). Glycerol, as a chemical chaperone, is a polyclonal T cell activator (Ghumman et al., 1998). CFA (a mix of TLR agonists) acts on TLRs as expressed in C57BL/6 mice resulting in both B and T cell activation.

The detected secretion of both Th1 (INF- γ , TNF- α , IL-12) and Th2 cytokines (IL-4 and IL-5) may be consequential to the influence of CFA upon T cells (Billiau & Matthys 2001). CFA recruits natural memory cells (MZ B and B1) to assist in the interrogation of B cells (Bernasconi et al., 2002, Ha et al., 2006). This in turn leads to the generation of an Ag-specific adaptive immune response based on archetypes contained in and drawn from the pool of all anti-bacterial memories; thus there would appear to be latent or innate blueprints which can be used to produce an interim specific Ab response, a stopgap to afford the breathing-space necessary to permit a conventional adaptive response to develop. The importance of TLR activation in the production of AAbs is implicit in the fact that it was the CFA-pretreated mice that produced AAbs more quickly and in greater quantities than did any other experimental group.

AgS may also be drivers of anti- β 2GPI and anti-ganglioside Ab production. TTd may incite anti- β 2GPI Ab production by molecular mimicry: TTd and β 2GPI share partial similarities of three-dimensional structure (Inic-Kanada et al., 2009). An alternative hypothesis is that anti-ganglioside Abs are generated in the context of Id/anti-Id networking; mice immunized (varying strategies) with gangliosides subsequently displayed poor antibody responses (Bowes et al., 2002). As the gangliosides G_{D1b} and G_{T1b} are tetanus toxin receptors, an antigen-binding site found on an anti-ganglioside IgG might be an internal image of an anti-TTd Ab, reactive towards the ganglioside-binding site found on the tetanus antigen. In an idiotypic network, anti-TTd IgG might be typed as Ab1 with anti-ganglioside Abs typed as Ab2. This hypothesis is not in conflict with the sequence of events observed during specific Ab secretion: an appreciable rise in anti-TTd IgG sera concentration precedes the secretion of anti-ganglioside IgG. The increase in IgG2c ganglioside-specific antibodies after the TTd booster dose carries with it the implication that both Id/anti-Id interactions and Th1 cytokines are together involved in the synthesis of these former.

If maintenance of tolerance to self-structures is paramount in the immune system hierarchy then protection against those microbial invaders which impersonate (to a greater or lesser extent) host self-structures is problematic; how to mount a defense against the invader without compromising this tolerance and inducing an autoimmune pathology? It seems inevitable that, in any rapid defense mounted, a tipping-point will be reached when polyclonal activation normally preventive (in a heuristically homeostatic system) of autoimmunity will itself initiate a runaway autoimmune response.

If polyclonal B cell activation alone (or changes in quantitative interactions in the public Id) acts as the trigger for autoimmunity then the regulatory potential of the NAAb network is nullified; the constitutive recurrence (dynamic and metadynamic properties of interactions) of the public Id

would be neglected and thus the full regulatory potential of NAAbs could never be exploited. Thus polyclonal B cell activation cannot be the sole culprit; somatic mutation and affinity maturation are prerequisites aided implicitly by regulator-T cells, helper-T and follicular B cells; a variety of antigens and strong autoantibodies are complicit.

It follows then that any ability to neutralize the threats posed by the multiplicity of near self-similar invasive bodies must be integral to a system in its entirety (reliant upon mutations in the public Id which whilst connected retains it utility) thus explaining the observed differences in the responses of BALB/c and C57BL/6 mice to TTd hyperimmunization.

Any heightened NAAb connectivity recruits autoreactive B cells to the network; these themselves then become subject to the dynamics of the network, both limiting their clonal reproductivity and inhibiting Ab secretion. These dynamics also shepherd NAAb complements into the network fold. The sequence then becomes recursively self-iterating, gathering in all functionally autoreactive B cells subsequently produced.

It is worth noting that, within newly produced or immunization-produced AAb pools, perturbances and synchronous fluctuations in AAb levels were induced; we were either witnessing a cross-reactivity with NAAbs or the AAbs were themselves members of a NAAb network. We saw that a quantitative disturbance in preexisting Id/anti-Id interactions was induced by TTd hyperimmunization and that this was most noticeable in groups pretreated with CFA or glyc.

As stated, if polyclonal B cell activation is the primary factor in the initiation and continued induction of autoimmunity in BALB/c mice, then the ability of C57BL/6 mice to sidestep the threats arising from TTd hyperimmunization must be genetically integral to their immune system. Let us now attempt to explore this issue further.

The question: why does an identical immunization protocol induce a characteristic pathology (APS with fetal resorption) in BALB/c mice yet not do so in C57BL/6 mice? Consider first the numerous V_H functional genes, differently expressed in different mouse strains. The relative map positions of the V_H gene families in Igha and Ighb haplotypes are indistinguishable (Brodeur et al., 1988) and it is thus unlikely that the differential expression of V_H families in BALB/c and C57BL/6 mice is due to differences in V_H gene organization. But it is documented that, in the C57BL/6 strain, anti-PtC Abs are mainly encoded by V_H1 immunoglobulin heavy-chain variable region gene families; usage of the V_H2 gene predominates in BALB/c mice. In the anti-PtC Ab, heavy-chains encoded by the V_H1 and V_H2 genes are combined with the V κ 9 and V κ 4 light-chain genes respectively; J_H1 genes combine with third complementary determining regions (CDR3s) of longitudinal homogeneity (Poncet et al., 1990, Carmack et al., 1990). Diversity within the anti-PtC Ab pools encoded by V_H1 is assured by the high number of genes within that family, greatly increasing the likelihood that a broad spectrum of germ-line encoded end products will be generated (Seidl et al., 1999).

AAbs encoded by the significantly smaller V_H2 family (V_HQ52 gene family) display a greater diversity (N-region additions and variable length CDR3s) and so may easily slip

through the regulatory net of natural Id/anti-Id interactions (Seidl et al., 1999). That synchronous fluctuations within newly produced or immunization-produced AAb pools were seen in C57BL/6 mice (with Id/anti-Id disturbance most noticeable in pretreated animals) suggests that it is intense polyclonal stimulation in a milieu of Id/anti-Id interactions that keeps the system in an organized state, with any detectable pathologies eliminated either through hysteresis or the total suppression of an autoimmune response.

Self-tolerance is the sine qua non of the immune system as we understand it. Each new immune response is deemed to be generated from and within a network of preexisting NAAb idiotypes. It follows that a tetanus toxoid-specific immune response will be generated within the interstices of Id/anti-Id interactions framing phospholipid-specific NAABs. Abs to Ags such as PtC must exist innately, a priori to any Ag first encounter; it is hard if not impossible to posit that somatic mutation and selection alone can create and account for specificities to Ags yet to be encountered. It follows that an imperfect or inchoate NAAB network, consequential to a genetically predetermined deficiency in the library of NAAB producing cells, may leave BALB/c mice vulnerable to APS (Avrameas et al., 2007). The more diverse palette of AAbs found in C57BL/6 mice has painted a network capable of preventing APS.

ACKNOWLEDGEMENTS

This research was supported by a grant from the Ministry of Science and Technological Development of Republic of Serbia (contract numbers 142020 and 172049).

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