

EFFECT OF NITROGEN SOURCE ON SOME RHIZOSPHERIC PROPERTIES AND PERSISTENCE OF MYCORRHIZAL FUNGAL PROPAGULES IN AN ANDISOL

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ABSTRACT

Nitrogen fertilization influences plant growth and rhizospheric properties, affecting the functionality and persistence of arbuscular mycorrhizal fungi (AMF). In order to analyze the effect of two N-sources (NH_4^+ and NO_3^-) on the persistence of AMF propagules (colonized root, mycelium and spores) and some rhizospheric parameters (pH, P available), a greenhouse experiment was carried out using two wheat (*Triticum aestivum* L.) cvs. Otto and Metrenco, which were grown in an Andisol. Plant biomass was determined at the dry grain stage (Zadocks 99, 150 days after sowing, DAS), the density of AMF propagules was determined three months later (240 DAS), and pH and available P were determined at both stages. The results showed that N-source and cultivar influenced most of the studied variables. The NO_3^- + Metrenco combination showed the highest values for biomass, pH, available P and AMF spores (150, 13, 5 and 375% more than NH_4^+ + Otto interaction, respectively; $p < 0.001$). On the other hand, close relationships were found between biomass production and density of AMF spores ($r^2 = 0.89$; $p < 0.001$), suggesting that this propagule is quantitatively affected by host plant biomass production. This propagule is probably also indirectly affected by rhizospheric conditions. This result is of a special agronomic and ecological interest in acidic soils destined to annual crops, in which nitrogen fertilization is a habitual practice, being the N-source very influential in yields.

Key words: acidic soils, available P, wheat cultivars, nitrogen fertilization, rhizospheric pH, *Triticum aestivum*.

INTRODUCTION

Nitrogen fertilization is a habitual practice used by farmers in intensive agriculture systems, as occurs in volcanic soils (Ultisols and Andisols) of southern Chile, which cover a surface area of about 5×10^4 km², dedicated mainly to cereal-legume rotation (Besoain, 1985). Frequently, these types of soils present fertility limitations due to low P availability associated with high levels of exchangeable Al and low pH values (Baligar and Fageria, 1997). Thus, the application of different N-sources (in particular NO_3^- and NH_4^+) is of special relevance, since they represent an important fraction of cations or anions absorbed and/or accumulated by plants, affecting directly soil pH due to the physiological extrusion of H^+ or OH^- by the roots (Stange *et al.*, 1995; Gerendás *et al.*, 1997). Particularly, NH_4^+ also strongly influences the reduction of soil pH through the generation of H^+ produced by the nitrification process (Campillo and Rodríguez, 1984; Martens, 2001). These rhizospheric changes will directly affect the solubility and mobility of several nutrients (Siqueira and Moreira, 1997) and the activity of diverse groups of microorganisms, including both symbiotic and free-living species (Jeffries and Barea, 2001).

Among the soil microbial groups that play a key role in the different rhizospheric processes, the arbuscular mycorrhizal fungi (AMF) (phylum Glomeromycota) are able to establish symbiotic associations known as arbuscular mycorrhizas (AM) with the roots of most terrestrial vascular plants (Smith and Read, 1997; Barea, 1998; Schüssler *et al.*, 2001). The main function of AM is nutritional, since the fungal hyphae act as a complementary root system, increasing the absorption area and allowing acquisition of nutrients with low

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mobility in the soil, or that are situated far from the root (Baligar and Fageria, 1997; Bago *et al.*, 2000; Clark and Zeto, 2000). These characteristics of mycorrhizal symbiosis present a special relevance in acidic soils, since it favors the root absorption of typically limited elements in this type of soil, particularly P, N and several microelements (Clark *et al.*, 1999; Clark and Zeto, 2000; Borie *et al.*, 2002). Different agronomic practices can affect AM functionality, as well as the density of spores, mycelia and colonized roots present in the soil. Among others, the crop system, the rotation design and the amount and type of fertilizer used are of particular relevance (Mendoza and Borie, 1998; Borie and Rubio, 1999; Jeffries and Barea, 2001).

In contrast, it had been demonstrated that extraradical AMF hyphae can uptake NH_4^+ as NO_3^- from the soil (Johansen, 1999; Bago *et al.*, 2001; Hawkins and George, 2001), in addition to significant amounts of N-organic and amino acids (Hawkins *et al.*, 2000); however, to date, only a high-affinity NH_4^+ transporter expressed in extraradical AMF hyphae has been characterized (López-Pedrosa *et al.*, 2006). Functional AM can generate important changes of pH in the mycorrhizosphere, as has been detected in various assays using *in vitro* culture systems (Bago *et al.*, 1996; Bago and Azcón-Aguilar, 1997). It has been observed that the development of mycelium in the presence of NO_3^- is associated with a pH increase, and the opposite occurs in the presence of NH_4^+ . No concordance exists with respect to the effect of fertilization with different N-sources on mycorrhizal plants; some studies suggest their preference for NO_3^- (Azcón *et al.*, 1992), while others suggest their preference for NH_4^+ (Cuenca and Azcón, 1994), with genotypical variations being able to exist within the same plant species, mainly in limited N environments (Nakamura *et al.*, 2002).

The aims of the present study were: i) to determine the effect of two N fertilizers, such as urea and sodium nitrate, and two commonly cropped wheat cultivars on the persistence of AMF propagules in an Andisol, and ii) to analyze the interactive effect of both factors on the changes of some soil chemical characteristics.

MATERIALS AND METHODS

Experimental design. A 2 x 2 full factorial randomized experiment design was used, including two wheat cultivars commonly used in the study zone

(‘Otto’ and ‘Metrenco’) and two N sources (N-NH_4^+ and N-NO_3^-). Each treatment combination had four replicates for each of two measurement stages.

Soil characteristics. The test soil used was collected from a 5 to 25 cm depth of an annual crop site in the commune of Vilcún (38°41' S; 72°24' W), Region of the Araucanía, Chile. Some characteristics of the soil, an Andisol of the Vilcún series (Entyc Dystrandep), are presented in Table 1. Soil was air-dried, sieved through a 5 mm mesh, and supplied with 0.06 g P kg^{-1} soil as Triple Superphosphate and 0.063 g K kg^{-1} soil as KCl in solution. One liter pots were filled with 800 g of soil.

Plant material. Wheat cvs. Otto and Metrenco were used as host plants. Seeds of each cultivar were surface sterilized with a 2% Cloramin-T ($\text{CH}_3\text{C}_6\text{H}_4\text{SO}_2\text{N}(\text{Cl})\text{Na} \times \text{H}_2\text{O}$) solution for 3 min and rinsed thoroughly with distilled water. Four seeds were germinated between wet tissue paper and planted 7 d later. Pots were thinned to two plants after establishment.

Growth conditions. Plants were grown under greenhouse conditions with temperatures ranging from 25 ± 3 °C day-time to 15 ± 3 °C night-time, a 16:8 h light:dark photoperiod and a relative humidity of 80-90%. A photosynthetic photon flux density of 0.4-0.5 $\text{mmol m}^{-2} \text{s}^{-1}$ was applied as supplementary light when necessary. Plants were irrigated manually with distilled water as needed during the experiment (judged by weighing the pots). Every 2 wk, 10 mL of a nutrient solution (Johnson *et al.*, 1996), without P and N, was added to each pot. The N was supplied twice, at establishment (30% total N in Zadocks 11

Table 1. Selected chemical and microbiological characteristics of the Vilcún series soil (Entyc Dystrandep) used in this study.

Soil characteristic	Value
Available P, mg kg^{-1}	4.00
pH (H_2O)	5.42
Soil organic matter, %	18.00
CEC, $\text{cmol}(+)\text{kg}^{-1}$	11.33
Al saturation, %	0.61
AMF spores 100 g^{-1} soil	406
Total hyphae density, m g^{-1} soil	8.50
Active hyphae density, m g^{-1} soil	4.34

CEC: cation exchange capacity; AMF: arbuscular mycorrhizal fungi.

stage) and at 6 wk of cultivation (70% total N in Zadocks 31 stage) (Zadocks *et al.*, 1974) to an equivalent of 0.126 g N kg⁻¹ soil, which represents a normal fertilization rate equivalent to 200 kg N ha⁻¹. In the NO₃⁻ treatments, NaNO₃ was used; whereas CO(NH₂)₂ was used in the NH₄⁺ treatments. In both cases, N was supplied in solution.

Harvest and analyses. Two harvest stages were considered. The first stage was at dry grain (150 days after sowing (DAS), at Zadocks 99 stage), and the second stage was at 240 DAS (3 months after grain harvest). Only soil and roots were present in the pots in the last stage, and they had been watered once a week since grain harvest. The harvested roots and shoots were dried at 65 °C for 48 h in a forced-air oven (Memmert GmbH + Co., Mod. UIM 400, Schwabach, Germany) and weighed. Before drying, 1 cm root fragments were separated and AM colonization was estimated by the method described by Giovanetti and Mosse (1980), after clearing and staining with Trypan blue (Phillips and Hayman, 1970). Root length was determined by the gridline intersect method (Tennant, 1975). The colonized root was determined at 150 and 240 DAS. The total extraradical mycorrhizal hyphae were determined by the method described in Borie *et al.* (2000), and the active hyphae were determined using dehydrogenase activity (Sylvia, 1988; Kabir *et al.*, 1997). Total and active hyphal density were quantified using the intersect gridline method (Newman, 1966). Mycorrhizal spores were separated from soil by wet sieving and decanting in a 70% w/v sucrose solution (Gerdemann and Nicholson, 1963); spores were then quantified under a stereoscopic microscope at 50X. Extraradical hyphae and mycorrhizal spores were determined at 240 DAS. Soil pH was measured in a soil:water mix (2:5), and available P in the soil by the method described by Olsen and Sommers (1982) after extraction with 0.5 M NaHCO₃ (pH 8.5). Soil pH and available P were determined at 150 and 240 DAS.

Statistical analyses. The main effects of wheat cultivar, N source and their interaction were tested by means of a two-way analysis of variance (ANOVA) using the general linear model procedures of the SPSS software, version 10.0 (SPSS Inc.) (Pérez, 2001). Means were compared by the orthogonal contrast test (Petersen, 1977). Data sets not meeting assumptions for ANOVA were transformed as required, but the results were presented in their original scale of measurement. The analysis of the relationship between different

variables was carried out by lineal correlation. Statistical significance was determined at $p < 0.05$.

RESULTS AND DISCUSSION

The wheat cultivars and the N-sources used as fertilizers influenced the behavior of most of the analyzed variables (Table 2). In general, variables significantly affected by one factor were also affected by the other, as well as by their interaction. The shoot and root biomass production was greater in Metrenco cultivar, in comparison to Otto cultivar (Figure 1). The NO₃⁻ use was related to a biomass increase in both cultivars, the Metrenco + NO₃⁻ treatment registering a shoot biomass 150% higher than the Otto + NH₄⁺ treatment. These differences could be related to different genotype and phenotypic characteristics of cultivars, as is the case of root length or biomass. Previous observations of Nakamura *et al.* (2002),

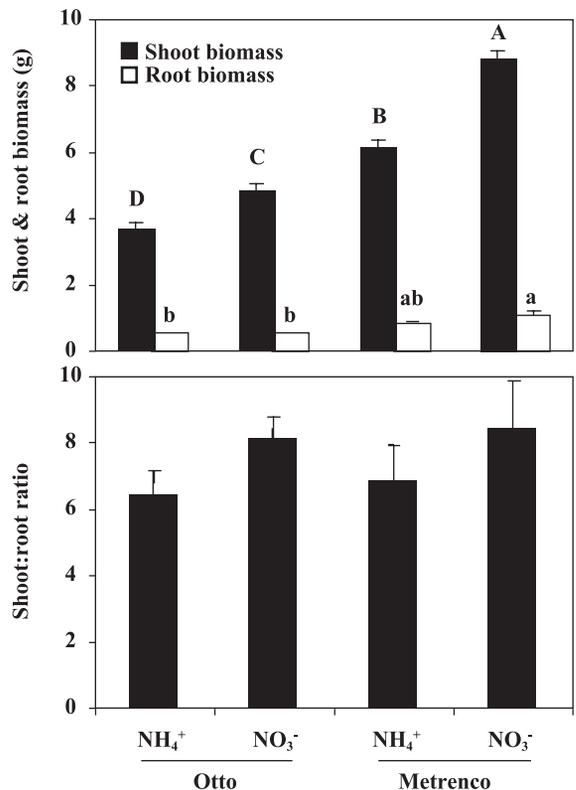


Figure 1. Biomass and shoot:root ratio of wheat plants cvs. Otto and Metrenco at 150 days after sowing fertilized with two N-sources (NO₃⁻ and NH₄⁺). The bars represent standard error. For each data series, bars with different letters indicate significantly different means ($p < 0.05$) using an orthogonal contrasts test.

Table 2. F-values and probabilities of significance for the main effects and factor interaction for the variables analyzed by means of a two-way ANOVA at 150 and 240 days after sowing (DAS).

	150 DAS			240 DAS		
	Cultivar	N source	C x N ¹	Cultivar	N source	C x N ¹
Shoot biomass, g	206.05***	75.06***	10.98**	-	-	-
Root biomass, g	14.32**	1.62 ^{ns}	1.19 ^{ns}	-	-	-
Shoot:root ratio	0.13 ^{ns}	2.35 ^{ns}	0.01 ^{ns}	-	-	-
Soil pH	189.70***	606.37***	18.60**	104.93***	132.80***	3.29 ^{ns}
Available P, mg kg ⁻¹	0.90 ^{ns}	0.09 ^{ns}	8.98*	0.71 ^{ns}	1.33 ^{ns}	1.86 ^{ns}
Total root length, m	3.10 ^{ns}	0.36 ^{ns}	2.59 ^{ns}	14.05**	10.34**	6.03*
AM root length, m	1.82 ^{ns}	0.03 ^{ns}	1.12 ^{ns}	13.44**	8.67**	5.87*
AM colonization, %	2.05 ^{ns}	1.22 ^{ns}	1.57 ^{ns}	1.12 ^{ns}	0.67 ^{ns}	0.83 ^{ns}
Total hyphae, m g ⁻¹ soil	-	-	-	0.20 ^{ns}	0.13 ^{ns}	4.49 ^{ns}
Active hyphae, m g ⁻¹ soil	-	-	-	2.75 ^{ns}	0.03 ^{ns}	4.15 ^{ns}
Hyphal activity, %	-	-	-	0.17 ^{ns}	0.01 ^{ns}	0.01 ^{ns}
AM fungi spores	-	-	-	32.60***	23.12***	5.78*

AM, arbuscular mycorrhizal; ns, not significant; * p < 0.05; ** p < 0.01; *** p < 0.001.

¹Interaction between the factors wheat cultivar (C) and N source.

working with native and hybrids sorghum (*Sorghum bicolor* (L.) Moench) cultivars, suggest that greater growth and radical activity are the main determining factors of higher biomass production by a particular cultivar. In the present study, the different cultivars presented differences in shoot biomass production (p < 0.01), independent of the N-source used, and similar shoot:root ratios in both cultivars were observed, although the use of NO₃⁻ was associated with moderate increases in this ratio (Figure 1).

Total and AM colonized root length did not present significant differences in any of the combinations studied at 150 DAS (Figure 2). However, it was observed that a greater amount of roots in cv. Metrenco (78.6% more than cv. Otto) at 240 DAS remained persistent, especially with NH₄⁺ use. In contrast, the mycorrhizal colonization was not affected by the analyzed factors in any of the studied stages, although an important increase of the mycorrhizal colonization at 240 DAS with regard to the previous stage was observed (Figure 2). The effect of seasonal changes in levels of colonization has been observed in other studies, although with different behavior. Thus, Kabir *et al.* (1997) found that higher levels of colonization appeared in summer, coinciding with more advanced development stages of maize (*Zea mays* L.) plants. On the other hand, Cornejo *et al.* (2007), in a greenhouse study carried out under similar conditions to the present ones, observed differences in levels of colonization only in early stages of a wheat crop; in

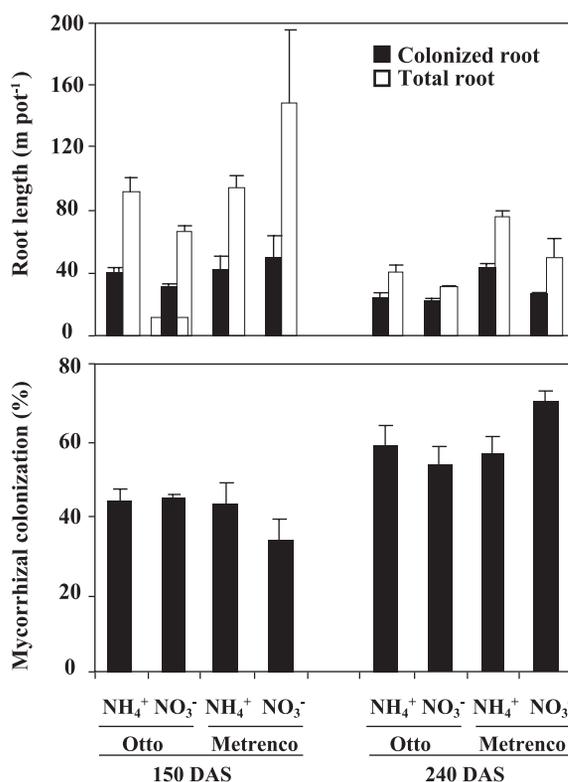


Figure 2. Total and colonized root length, and mycorrhizal colonization of wheat plants cvs. Otto and Metrenco at 150 and 240 days after sowing (DAS) fertilized with two N-sources (NO₃⁻ and NH₄⁺). The bars represent standard error.

addition, colonization was found to be greater when NO_3^- was used. As in the present study, colonization levels increased after plant harvest, which probably does not indicate new colonization, but rather a greater prevalence of the mycorrhizal roots at the end of the plant life cycle. Therefore, we can suppose that differences in the mycorrhizal colonization were present in previous growth stages, not considered in the present study. However, the high prevalence of mycorrhizal roots is an aspect of special agro-ecological relevance, since the mycorrhizal root fragment pieces are an important AMF inoculum source for the succeeding crop in the rotation (Brundrett, 1991).

A similar behavior was observed in the case of mycorrhizal colonization and persistent hyphae in the soil, and no significant differences between different combinations of cultivars and N-source were observed, nor were the total and active hyphae density or proportion of active hyphae (Figure 3). In a similar way, the study carried out by Cornejo *et al.* (2007) found, in a wheat crop, higher densities of active hyphae in NO_3^- fed soils only at dry grain harvest. This behavior could be determined by the AMF life cycle, a basal or residual hyphal density appearing in the first plant and symbiosis development stage, later being dependant on the intraradical colonization extension, finally becoming stabilized when the bidirectional nutrient exchange between plant and fungus stops because of vegetal senescence or, as in this study, because of the shoot harvest. In contrast,

densities of persistent hyphae were 2-3 folds higher in comparison to those of the Cornejo *et al.* (2007) report, probably because of the contribution to this component of several AMF species native to the natural soil. Pringle and Bever (2002) found that the AMF community composition was modified over time due to the proliferation of different fungi species, independently of other factors analyzed, such as the floristic community composition. It is possible that the adaptation to specific situations by some AMF species, as in this case of different N-sources, the wheat cultivar used, or the different conditions and variations of soil characteristics, confers a high plasticity to the natural AMF communities, allowing them to reach similar densities of fungal propagules by the contribution of different species. Similarly, Munkvold *et al.* (2004) found a high intraspecific diversity in the mycelial growth of different AMF species, suggesting the existence of an important functional complementariness, including at low levels of fungal diversity. Thus, studies of diversity and population dynamics that present different species within the AMF communities become quite necessary.

The fungus life cycle is completed with the production of resistance spores (Brundrett, 1991), which are able to generate new colonization. In this study, highly significant differences in the AMF spore density between the different treatments, attributable to both studied factors, were registered (Figure 4). ‘Metrenco’ presented higher spore density than ‘Otto’; in addition, the use of NO_3^- as N-source favored the formation of

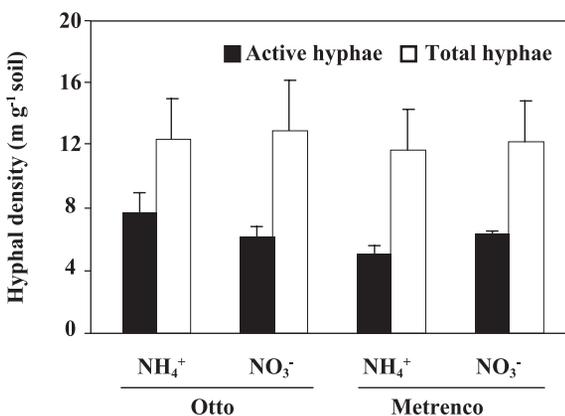


Figure 3. Density of active and total persistent extraradical mycorrhizal hyphae at 240 days after sowing in two wheat cultivars (Otto and Metrenco) fertilized with two N-sources (NO_3^- and NH_4^+). The bars represent standard error.

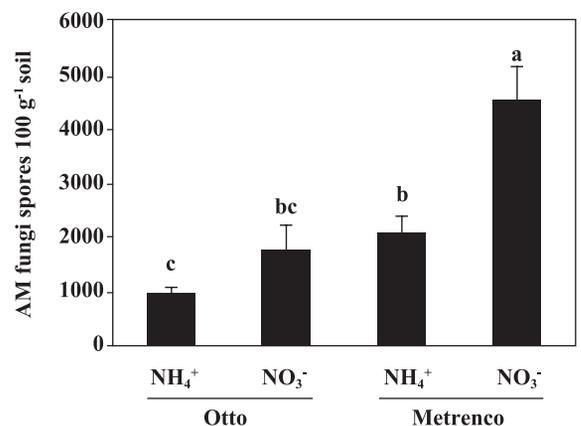


Figure 4. Persistence of arbuscular mycorrhizal (AM) fungi spores at 240 days after sowing in two wheat cultivars (Otto and Metrenco) fertilized with two N-sources (NO_3^- and NH_4^+). The bars represent standard error.

spores in both cultivars. The Metrenco + NO₃⁻ treatment produced 375% more spores than did the combination Otto + NH₄⁺. No clear relationship between the persistent spore density and some other AMF propagule was found. However, shoot biomass production was high and directly related to spore production ($r^2 = 0.89$; $p < 0.001$). This result could be based on the greater carbonated flux than presumably was obtained in plants with a greater growth, allowing a greater carbonated compounds contribution to intraradical fungal mycelium, which by glyconeogenesis in external mycelium would later be transformed into reserve lipids (Pfeffer *et al.*, 1999; Bago *et al.*, 2002), an important component of resistance spores.

The results presented by Cornejo *et al.* (2007) suggest greater spore production in the final growth stages in annual crops, which could be based on the functionality of other AM components in previous stages of the fungal life cycle, such as intraradical colonization and subsequent soil colonization by extraradical hyphae. These results and observations are of significant agronomic and ecological interest, since a greater mycorrhizal propagule density will lead to a greater probability of generating new and early-effective fungal colonization in the succeeding crop in the rotation.

In the case of soil-available P, it was observed that the Metrenco + NO₃⁻ combination had significant

differences only with regard to the Otto + NO₃⁻ combination (3 mg P kg⁻¹ of difference between both) (Table 3). In particular, cv. Otto exhibited a significant difference between the N-sources used, the NH₄⁺ use being associated with greater soil P availability. Previous studies have also reported greater P availability in soils fertilized with NH₄⁺ sources relative to NO₃⁻ sources (Ortas *et al.*, 1996). In the present study, the increase in P availability was associated with a greater colonized root length in 'Otto' at 150 DAS; a greater functionality of the established mycorrhiza could explain this greater P availability. However, Ortas *et al.* (1996) suggest that a greater P availability can result from a stimulating effect of NH₄⁺ on the roots, producing a concomitantly greater organic acid exudation that would release P from different soil compartments, a mechanism that has been widely studied (Hoffland *et al.*, 1989; Johnson *et al.*, 1996; Montenegro and Zapata, 2002). Of the soil-root interface properties analyzed, only pH exhibited important differences by effect of the studied factors, these differences being higher at 150 DAS (Table 2 and 3). The pH values were greater in 'Metrenco', especially when NO₃⁻ was used; a difference of 0.69 units of pH at 150 DAS was measured with regard to the Otto + NH₄⁺ combination. This difference persisted at 240 DAS. These results agree with previous observations demonstrating that the rhizospheric pH changes are due to the N-source used and to the mycorrhizal plant status (Vaast and Zasoski, 1992; Bago and Azcón-Aguilar, 1997).

Table 3. Soil-root interface traits at 150 and 240 days after sowing (DAS) in two wheat cultivars (Otto and Metrenco) fertilized with two N-sources (NO₃⁻ and NH₄⁺).

Cultivar	N source	150 DAS	240 DAS
		Available P (mg kg ⁻¹)	
Otto	NH ₄ ⁺	9.27a	8.13a
	NO ₃ ⁻	6.75b	7.98a
Metrenco	NH ₄ ⁺	7.70ab	6.58a
	NO ₃ ⁻	9.77a	8.34a
pH (H₂O)			
Otto	NH ₄ ⁺	5.35d	5.18b
	NO ₃ ⁻	5.72b	5.49ab
Metrenco	NH ₄ ⁺	5.52c	5.46ab
	NO ₃ ⁻	6.04a	5.69a

For each cultivation stage, means followed by the same letter in a column are not significantly different using orthogonal contrasts test ($p < 0.05$; $n = 4$).

The observed alkalization due to NO_3^- absorption by the roots (and the mycorrhiza) is well-known (Marschner, 1995; Bago and Azcón-Aguilar, 1997). Nevertheless, the effect that different wheat cultivars exhibited on pH can be explained by different biomass production. The greater growth achieved by 'Metrenco' could have made possible a greater absorption and a faster assimilation of NH_4^+ , avoiding its use as a substrate in the nitrification processes that, as has been observed in volcanic soils in southern Chile, are able to induce strong pH reductions (Campillo and Rodríguez, 1984). In the present study, highly significant correlations between pH values and shoot biomass production ($r^2 = 0.83$ at 150 DAS; $p < 0.001$) were found, which could reinforce the previous observation. As well, a highly significant correlation between pH values and AMF spore production at 240 DAS ($r^2 = 0.80$; $p < 0.001$) was found, although this could reflect an indirect effect of a greater biomass production of the Metrenco + NO_3^- treatment and a greater contribution of carbonated compounds than a direct influence of the soil acidity.

CONCLUSIONS

The use of best adapted cultivars and/or those with a greater biomass production potential, as was observed in the case of Metrenco, can be associated with an increase in the mycorrhizal functionality, which will produce an increase in propagule density. On the other hand, the application of NO_3^- favored plant growth, which was related to a significant decrease of soil acidity, one of the commonest limiting characteristics of volcanic soils such as that used in this study. The improvement of soil characteristics using NO_3^- was directly related to higher persistence of AMF spores in the soil, even at similar levels of hyphal length and colonized roots, demonstrating its strong impact on the native AMF populations present in Andisols in southern Chile. In summary, the use of suitable agricultural practices allows greater plant biomass production, an important improvement of soil chemical conditions and superior persistence of mycorrhizal propagules, particularly spores, which will produce earlier effective plant colonization in the succeeding crop in the rotation. This last aspect is very important from a realistic point of view, considering that these kinds of soils are used mainly in annual rotations, providing supplementary tools to farmers for a better choice of N-source.

R E S U M E N

Efecto de la fuente de nitrógeno sobre algunas propiedades rizosféricas y persistencia de propágulos de hongos micorrícicos en un Andisol. Pablo Cornejo^{1*}, Rosa Rubio¹ and Fernando Borie¹. La fertilización nitrogenada influye en el crecimiento vegetal y propiedades rizosféricas, afectando la funcionalidad y persistencia de hongos micorrícico-arbusculares (AMF). Para analizar el efecto de dos fuentes de N (NH_4^+ y NO_3^-) sobre la persistencia de propágulos de AMF (raíz colonizada, esporas e hifas) y algunos parámetros rizosféricos (pH, P disponible), se realizó un experimento en invernadero utilizando dos cultivares de trigo (*Triticum aestivum* L.) cvs. Otto y Metrenco en un Andisol. La biomasa se determinó en grano seco (Zadocks 99, 150 días después de la siembra, DAS), la densidad de propágulos de AMF se determinó tres meses más tarde (240 DAS), y el pH y P disponible se determinaron en ambas etapas. Los resultados indicaron que la fuente de N y el cultivar influyeron en la mayoría de las variables estudiadas. La combinación NO_3^- + Metrenco presentó mayores valores de biomasa, pH, P disponible y esporas de AMF (150; 13; 5 y 375% más que la interacción NH_4^+ + Otto, respectivamente; $p < 0,001$). Por otra parte, se encontraron estrechas relaciones entre producción de biomasa y densidad de esporas de AMF ($r^2 = 0,89$; $p < 0,001$), que sugiere que este propágulo es cuantitativamente afectado por la producción de biomasa de la planta hospedera. Este propágulo es probablemente afectado de forma indirecta por las condiciones rizosféricas. Este resultado es de especial interés ecológico y agronómico en suelos ácidos destinados a cultivos anuales, donde la fertilización nitrogenada es una práctica habitual, siendo la fuente de N muy incidente en los rendimientos.

Palabras clave: suelos ácidos, P disponible, cultivares de trigo, fertilización nitrogenada, pH rizosférico *Triticum aestivum*.

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LITERATURE CITED

- Azcón, R., M. Gómez, and R. Tobar. 1992. Effects of nitrogen source on growth, nutrition, photosynthetic rate and nitrogen metabolism of mycorrhizal and phosphorus-fertilized plants of *Lactuca sativa* L. *New Phytol.* 121:227-234.
- Bago, B., and C. Azcón-Aguilar. 1997. Changes in the rhizospheric pH induced by arbuscular mycorrhiza formation in onion (*Allium cepa* L.). *Z. Pflanzenernah. Bodenk.* 160:333-339.
- Bago, B., C. Azcón-Aguilar, Y. Shachar-Hill, and P. Pfeffer. 2000. El micelio externo de la micorriza arbuscular como puente simbiótico entre la raíz y su entorno. p. 78-92. *In* Alarcón, A., and R. Ferrera-Cerrato (eds.) *Ecología, fisiología y biotecnología de la micorriza arbuscular*. Colegio de Postgraduados, Ediciones Mundi Prensa, Montecillo, México.
- Bago, B., P. Pfeffer, and Y. Shachar-Hill. 2001. Could the urea cycle be translocating nitrogen in the arbuscular mycorrhizal symbiosis? *New Phytol.* 149:4-8.
- Bago, B., P. Pfeffer, W. Zipfel, P. Lammers, and Y. Shachar-Hill. 2002. Tracking metabolism and imaging transport in arbuscular mycorrhizal fungi. *Plant Soil* 244:189-197.
- Bago, B., H. Vierheilig, Y. Piché, and C. Azcón-Aguilar. 1996. Nitrate depletion and pH changes induced by the extraradical mycelium of the arbuscular mycorrhizal fungus *Glomus intraradices* grown in monoxenic culture. *New Phytol.* 13:273-280.
- Baligar, V.C., and N.K. Fageria. 1997. Nutrient use efficiency in acid soils: nutrient management and plant use efficiency. p. 75-95. *In* Moniz, A., A. Furlani, R. Schaffert, N. Fageria, C. Rosolem, and H. Cantarella (eds.) *Plant-soil interactions at low pH: Sustainable agriculture and forestry production*. Brazilian Soil Science Society, Campinas, São Paulo, Brazil.
- Barea, J.M. 1998. *Biología de la rizósfera*. *Investigación y Ciencia* 256:74-81.
- Besoain, E. 1985. Los suelos. p. 23-106. *In* Tosso, J. (ed.) *Suelos volcánicos de Chile*. Instituto de Investigaciones Agropecuarias, Santiago, Chile.
- Borie, F., Y. Redel, R. Rubio, J.L. Rouanet, and J.M. Barea. 2002. Interactions between crop residues application and mycorrhizal developments and some soil-root interface properties and mineral acquisition by plants in an acidic soil. *Biol. Fertil. Soils* 36:151-160.
- Borie, F., and R. Rubio. 1999. Effects of arbuscular mycorrhizae and liming on growth and mineral acquisition of aluminium-tolerant and aluminium-sensitive barley cultivars. *J. Plant Nutr.* 22:121-137.
- Borie, F., R. Rubio, A. Morales, and C. Castillo. 2000. Relationships between arbuscular mycorrhizal hyphal density and glomalin production with physical and chemical characteristics of soil under no-tillage. *Rev. Chil. Hist. Nat.* 73:749-756.
- Brundrett, M. 1991. Mycorrhizas in natural ecosystems. *Adv. Ecol. Res.* 21:171-291.
- Campillo, R., and J. Rodríguez. 1984. Efecto acidificante de las transformaciones de la urea en dos Andisoles de la región de Los Lagos. *Agric. Téc. (Chile)* 44:131-138.
- Clark, R.B., and S.K. Zeto. 2000. Mineral acquisition by arbuscular mycorrhizal plants. *J. Plant Nutr.* 23:867-902.
- Clark, R.B., S.K. Zeto, and R.W. Zobel. 1999. Arbuscular mycorrhizal fungal isolate effectiveness on growth and root colonization of *Panicum virgatum* in acidic soil. *Soil Biol. Biochem.* 31:1757-1763.
- Cornejo, P., F. Borie, R. Rubio, and R. Azcón. 2007. Influence of nitrogen source on the viability, functionality and persistence of *Glomus etunicatum* fungal propagules in an Andisol. *Appl. Soil Ecol.* 35:423-431.
- Cuenca, G., and R. Azcón. 1994. Effects of ammonium and nitrate on the growth of vesicular-arbuscular mycorrhizal *Eiyythrina poeppigiana* O.I. cook seedlings. *Biol. Fertil. Soils* 18:249-254.
- Gerdemann, J., and T. Nicholson. 1963. Spores of mycorrhizal endogone species extracted from soil by wet sieving. *Trans. Brit. Mycol. Soc.* 46:234-235.
- Gerendás, J., Z. Zhu, R. Bendixen, R. Ratcliffe, and B. Sattelmacher. 1997. Physiological and biochemical processes related to ammonium toxicity in higher plants. *Z. Pflanzenernah. Bodenk.* 160:239-251.
- Giovanetti, M., and B. Mosse. 1980. An evaluation of techniques for measuring vesicular-arbuscular mycorrhizal infection in roots. *New Phytol.* 84:489-500.
- Hawkins, H.J., and E. George. 2001. Reduced ¹⁵N-nitrogen transport through arbuscular mycorrhizal hyphae to *Triticum aestivum* L. supplied with ammonium vs. nitrate nutrition. *Ann. Bot.* 87:302-311.
- Hawkins, H.J., A. Johansen, and E. George. 2000. Uptake and transport of organic and inorganic nitrogen by arbuscular mycorrhizal fungi. *Plant Soil* 226:275-285.
- Hoffland, E., G.R. Findenegg, and J.A. Nelemans. 1989. Solubilization of rock phosphate by rape. 2. Local root exudation of organic-acids as a response to P-starvation. *Plant Soil* 113:161-165.

- Jeffries, P., and J.M. Barea. 2001. Arbuscular mycorrhiza - a key component of sustainable plant-soil ecosystems. p. 95-113. *In* Hock, B. (ed.) The mycota. Volume IX. Fungal associations. Springer-Verlag, Berlin, Germany.
- Johansen, A. 1999. Depletion of soil mineral N by roots of *Cucumis sativus* L. colonized or not by arbuscular mycorrhizal fungi. *Plant Soil* 209:119-127.
- Johnson, J.F., C.P. Vance, and D.L. Allan. 1996. Phosphorus deficiency in *Lupinus albus*. Altered lateral root development and enhanced expression of phosphoenolpyruvate carboxylase. *Plant Physiol.* 112:31-41.
- Kabir, Z., I.P. O'Halloran, J.W. Fyles, and C. Hamel. 1997. Seasonal changes of arbuscular mycorrhizal fungi as affected by tillage practices and fertilization: Hyphal density and mycorrhizal root colonization. *Plant Soil* 192:285-293.
- López-Pedrosa, A., M. González-Guerrero, A. Valderas, C. Azcón-Aguilar, and N. Ferrol. 2006. *Gint*AMT1 encodes a functional high-affinity ammonium transporter that is expressed in the extraradical mycelium of *Glomus intraradices*. *Fungal Genet. Biol.* 43:102-110.
- Marschner, H. 1995. Mineral nutrition of higher plants. 2nd ed. Academic Press, London, UK.
- Martens, D. 2001. Nitrogen cycling under different soil management systems. *Adv. Agron.* 70:143-189.
- Mendoza, J., and F. Borie. 1998. The effects of *Glomus etunicatum* inoculation on aluminium, phosphorus, calcium and magnesium uptake in two barley genotypes with different aluminium tolerance. *Commun. Soil Sci. Plant Anal.* 9:681-695.
- Montenegro, A., and F. Zapata. 2002. Rape genotypic differences in P uptake and utilization from phosphate rocks in an Andisol of Chile. *Nutr. Cycl. Agroecosys.* 63:27-33.
- Munkvold, L., R. Kjoller, M. Vestberg, S. Rosendahl, and I. Jakobsen. 2004. High functional diversity within arbuscular mycorrhizal fungi. *New Phytol.* 164:357-364.
- Nakamura, T., J.J. Adu-Gyamfi, A. Yamamoto, S. Ishikawa, H. Nakano, and O. Ito. 2002. Varietal differences in root growth as related to nitrogen uptake by sorghum plants in low-nitrogen environment. *Plant Soil* 245:17-24.
- Newman, E. 1966. A method of estimating the total length of root sample. *J. Appl. Ecol.* 3:139-145.
- Olsen, S.R., and L.E. Sommers. 1982. Phosphorus. p. 403-430. *In* A.L. Page, R.H. Miller, and D.R. Keeney (eds.) Methods of soil analysis. Part 2. Chemical and microbiological properties. Agronomy Monograph N° 9 American Society of Agronomy and Soil Science of America, Madison, Wisconsin, USA.
- Ortas, I., P.J. Harris, and D.L. Rowell. 1996. Enhanced uptake of phosphorus by mycorrhizal sorghum plants as influenced by forms of nitrogen. *Plant Soil* 184:255-264.
- Pérez, C. 2001. Técnicas estadísticas con SPSS. Prentice Hall, Madrid, España.
- Petersen, R. 1977. Use and misuse of multiple comparison procedures. *Agron. J.* 69:205-208.
- Pfeffer, P.E., D.D. Douds, G. Bécard, and Y. Shachar-Hill. 1999. Carbon uptake and the metabolism and transport of lipids in arbuscular mycorrhiza. *Plant Physiol.* 120:587-598.
- Phillips, J.M., and D.S. Hayman. 1970. Improved procedure for clearing roots and staining parasitic and vesicular-arbuscular mycorrhizal fungi for rapid assessment of infection. *Trans. Br. Mycol. Soc.* 55:158-161.
- Pringle, A., and J.D. Bever. 2002. Divergent phenologies may facilitate the coexistence of arbuscular mycorrhizal fungi in a North Carolina grassland. *Am. J. Bot.* 89:1439-1446.
- Schüssler, A., D. Schwarzott, and C. Walker, C. 2001. A new fungal phylum, the Glomeromycota: phylogeny and evolution. *Mycol. Res.* 103:1413-1421.
- Siqueira, J., and F. Moreira. 1997. Microbial populations and activities in highly-weathered acidic soils: highlights of the Brazilian research. p. 139-156. *In* Moniz, A., A. Furlani, R. Schaffert, N. Fageria, C. Rosolem, and H. Cantarella (eds.). Plant-soil interactions at low pH: Sustainable agriculture and forestry production. Brazilian Soil Science Society, Campinas, São Paulo, Brazil.
- Smith, S.E., and D.J. Read. 1997. Mycorrhizal symbiosis. Academic Press, London, UK.
- Stange, B., E. Beratto, A. Montenegro, A. Peyrelongue, and F. Borie. 1995. Effect of nitrogen source on barley growing in a high aluminium content soil. *Agric. Téc. (Chile)* 55:118-126.
- Sylvia, D. 1988. Activity of external hyphae of vesicular-arbuscular mycorrhizal fungi. *Soil Biol. Biochem.* 20:39-40.
- Tennant, D. 1975. A test of a modified line intersect method of measuring root length. *J. Ecol.* 63:995-1001.
- Vaast, P., and R.J. Zasoski. 1992. Effects of VA-mycorrhizae and nitrogen sources on rhizosphere soil characteristics, growth and nutrient acquisition of coffee seedlings (*Coffea arabica* L.). *Plant Soil* 147:31-39.
- Zadocks, J.C., T.T. Chang, and C.F. Konzak. 1974. A decimal code for the growth stages of cereals. *Weed Res.* 14:415-421.