USING AN INDOL DERIVATIVE AS COMPLEXING AGENT FOR CLOUD POINT PRECONCENTRATION AND DETERMINATION OF MAGNESIUM AND SILVER IONS IN VARIOUS SAMPLES BY FAAS

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ABSTRACT

A very simple cloud point extraction (CPE) methodology for the preconcentration of trace amounts of silver and magnesium as a prior step for their determination by flame atomic absorption spectrometry (FAAS) has been developed. The method is based on the extraction of silver and magnesium at pH 10 by using non-ionic surfactant Triton X-114 and 4 methyl-3-((1-H-Indol-3-iyl) (phenyl) methyl))-1-H-Indol (MPBIM) as a complexing agent. Several important variables that affect the CPE efficiency and FAAS signal were investigated and optimized. The calibration curves, using the preconcentration system, were linear in the range of 0.007–0.200 μg mL−1 with a correlation coefficient of 0.999. The lower limits of detection (3σ) obtained in the optimal conditions were 1.47 ng mL−1 and 3.00 ng mL−1 for Mg2+ and Ag+ ions, respectively. The relative standard deviation for five replicate determinations of magnesium and silver at 0.133 μg mL−1 concentration level, were 0.17 and 0.97, respectively. The proposed method was successfully applied to the ultra-trace determination of silver and magnesium in various water samples, blood serum, urine, and radiology film samples.

Keywords: Silver; Magnesium; Cloud-point extraction; Flame atomic absorption

INTRODUCTION

The determination of alkaline earth metals, particularly magnesium is of importance in environmental, biological and industrial applications. Magnesium, as a physiologically essential element, plays a significant role in the quality of drinking water.1 The relationship between magnesium deficiency and diabetes mellitus has been controversial in the medical field in recent years.2 In addition, Mg (II) is involved in a series of metabolism processes in the human body, including the formation of bones and cells.3 Widespread application of silver in industry, foods, drugs, beverages, photographic film production and the manufacturing of fungicides4,5 have resulted in increasing silver content of environmental water samples. In turn, owing to the toxicity of silver for many aquatic organisms even at low concentrations, serious environmental problems may occur. Therefore, simple and highly sensitive methods are required to monitor the Ag levels in water samples at ever decreasing concentrations.

A variety of methods including flame atomic absorption spectrometry (FAAS),6–8 electrothermal atomic absorption spectrometry (ETAAS),6,9 inductively coupled plasma optical emission spectrometry (ICP-OES),10,11 inductively coupled plasma mass spectrometry (ICP-MS)12–15 and stripping voltammetry16 have extensively been used for trace element determination in different samples. Direct determination of trace amounts of metal ions in some samples by flame atomic absorption spectrometry is difficult because of low sensitivity. Thus, pre-concentration procedures are often required. Different techniques, such as solvent extraction,15,16 membrane filtration,17 cloud point extraction18,19 and solid phase extraction20,21 have been used to enrich the analyte and separate it from the interferences.

The principal advantages and applications of CPE have been well-established and identified in recent years. The use of preconcentration steps based on CPE offers a conventional alternative to more traditional extraction systems and permits the design of extraction schemes that are simple, cheap, of high efficiency and of lower toxicity than extractions that use organic solvents. To date, CPE has been used for the extraction and preconcentration of metal ions after the formation of sparingly water-soluble complexes; then the complex in the surfactant-rich phase is determined by different spectrometric methods.22–24 In the present work, we report on the results obtained from applying the CPE for the extraction of Ag+ and Mg2+ ions from various samples and their determination by flame atomic absorption spectrometry.

EXPERIMENT AND PROCEDURE

Reagents
All solutions were prepared with double distilled water. Analytical-grade methanol, acids, and other chemicals were used (Merck, Darmstadt, Germany). A 1.0% (w/v) Triton X-114 from E. Merck was prepared by dissolving 1.0 g of Triton X-114 in 100 mL volumetric flask with stirring. All chemicals such as nitrate of Mg(II) and Ag(I) and other cations were the analytical grade purchased from Merck Company. 4 methyl-3-((1-H-Indol-3-iyl) (phenyl) methyl))-1-H-Indol (MPBIM) (Fig. 1) was synthesized according to the literature.22

Fig. 1. Structure of 4 methyl-3-((1-H-Indol-3-iyl) (phenyl) methyl))-1-H-Indol (MPBIM).

Instrumentation
A Sens. AA GBC (Australian) double beam atomic absorption spectrometer equipped with deuterium background correction and silver and magnesium hollow cathode lamps as the radiation source were used for absorbance measurements at wavelengths of 338 and 202 nm, respectively. The acetylene flow rate and burner height were adjusted in order to obtain the maximum absorbance signal, while aspirating the analyte solution in methanolic nitric acid. A Fater Ltd. model B 610 W thermostated water bath was employed to maintain the experimental temperature and a Shimifan model CE 148 centrifuge was used to accelerate the phase separation process. A Jenway pH meter model 3510 was used for pH measurements. An Adam model 220 LA electronic analytical balance was used for weighing the solid materials.

Procedure for cloud point extraction

A typical cloud point experiment required the following steps: 2 mL of a stock solution containing Mg\(^{2+}\) (1.0 μg mL\(^{-1}\)) and Ag\(^{+}\) (1.0 μg mL\(^{-1}\)) ions, 1 mL of Triton X-114 (1% w/v) and 1.5 mL MPBIM (1 mM) was placed into centrifuge tube and adjusted to pH 10 by the addition of NaOH and the total volume was reached to 15 ml in a marked centrifuge tube with double distilled water. The concentration of analytes in the prepared solution was 0.133 μg mL\(^{-1}\). The mixture was shaken for 1 min and left to stand in a thermostated bath at 65 °C for 20 min. The separation of the phases was achieved by centrifugation at 4000 rpm for 15 min. The whole system was cooled in an ice bath for 5 min so that the surfactant rich phase would regain its viscosity. In this way, the bulk aqueous phase was easily decanted. The remaining micellar phase was dissolved to 0.5 ml in tetrahydrofuran (THF)\(^{34}\) and the Mg\(^{2+}\) and Ag\(^{+}\) ions content were readily evaluated by FAAS.

Sample preparation
A: Water samples

Analysis of water samples for the determination of analyte contents was performed in the following way: About 500 mL of sample was passed through a sintered glass funnel and 1 mL concentrated HNO\(_3\) was added for elimination and decomposition of organic matter. The water samples were stored in polyethylene bottles. Then procedure given in general procedure section was applied for analysis.

B: Blood serum sample

A 5-mL portion of the clear supernatant serum obtained from a female with 34 years of age was pipetted off and treated with 5.0 mL of 1.2 M trichloroacetic acid. The mixture was centrifuged at 3000 rpm, and the deproteinizing supernatant was placed in a 50-mL beaker, and the resulting solution was neutralized with 2.0 M NaOH.\(^{31}\) Aliquots (7-mL) of the samples were then analyzed by the proposed method.

C: Urine sample

A 10 mL portion of a urine sample (or a spiked urine sample) was treated with 10 mL of concentrated HNO\(_3\) (63%) and HClO\(_3\) (70%) mixture of 2:1 in a 50 mL beaker covered with a watch glass. The content of the beaker was heated on a hot plate (100 °C 15 min, 150 °C 10 min). The watch glass was removed and the acid evaporated to dryness at 150 °C. HClO\(_3\) (3 ml) was added to the resulting white residue and the mixture was heated at 160 °C to dryness. All heating was carried out under a hood with necessary precautions. Five milliliters of 1 M H\(_2\)SO\(_4\) was added and the mixture was heated at 150 °C for 1 min. The neutralization was achieved by NaOH and the volume was made up to the mark in a 50 mL volumetric flask.\(^{34}\) Aliquots (7 mL) of the resulting clear solution were analyzed according to the described procedure.

D: Radiology film sample

The waste radiographic samples were prepared for the measurement of their silver content as follows. To 20 mL of the sample was added 10 mL 3 M nitric acid and 40 mL water. The solution was boiled for 60 min until its volume was reduced to a half. The resulting solution was neutralized with sodium hydroxide solution and subsequently filtered. The filtrate and washings were diluted to 100 mL in a volumetric flask and 2 mL of it, used for cloud point extraction and atomic absorption measurements.

RESULTS AND DISCUSSION

The aim of this paper is to develop a simple, sensitive and available method for the preconcentration and determination of trace amounts of Mg\(^{2+}\) and Ag\(^{+}\) ions in various real samples, using flame atomic absorption spectrometry coupling with CPE. In this regard, the influence of various effective parameters including pH, surfactant and ligand concentrations, heating time and temperature, centrifuge time and rate and the effect of electrolyte on absorbance were optimized.

Effect of pH

The formation of the metal–chelate and its chemical stability are two important factors influencing CPE. The pH plays a unique role in metal–chelate formation and subsequent extraction and proved to be the main parameter for CPE. Extraction yield depends on the pH at which complex formation is carried out. In this view, a set of similar experiments in the pH range of 2.0-11.0 was conducted according to the described procedure in the experimental section and the respective results are illustrated in Fig. 2. The maximum sensitivity by CPE was obtained at pH 10. In more acidic solutions, deterioration of the signal occurs due to the protonation of ligand, while at pH > 10, the signal decreases and the recovery is due to precipitation of Mg\(^{2+}\) and Ag\(^{+}\) ions in the form of hydroxides or ternary complexes. Also in strong alkali environments, the Triton X-114 was depersonated and the charge of surfactant prevents the formation of cloud. Consequently, pH = 10 was selected for the subsequent studies.
physicochemical characteristics, low cloud point temperature, commercial availability, relatively low price, low toxicity, its high density in the surfactant-rich phase and facility for phase separation. The variation in absorbance of extracted Mg\(^{2+}\) and Ag\(^{+}\) ions within the Triton X-114 concentration range of 7x10\(^{-3}\)–0.167 % (w/v) was examined and the results are shown in Fig. 4. It can be seen that quantitative extraction was obtained with an optimum Triton X-114 concentration of 0.06% (w/v), where the highest absorbance for extracted Mg\(^{2+}\) and Ag\(^{+}\) ions was obtained. For lower than 0.06% (w/v), the preconcentration efficiency of the formed complexes was very low, since the assemblies at low concentration were probably inadequate to preconcentrate trace amounts of Mg\(^{2+}\) and Ag\(^{+}\) ions. The reduction of absorbance in higher concentration than 0.06% (w/v), is due to the remaining of some part of Triton X-114 and ligand in aqueous solution as this phase can be competitive with surfactant-rich phase for drawing analyte ions.

![Fig.4. Effect of Triton X-114 concentration on the CPE of magnesium and silver ions. Conditions: Mg\(^{2+}\), 0.133 M Ag\(^{+}\), 0.133 M; Ligand, 0.1 mM; pH, 10.0](image)

**Effect of ionic strength**

It is known that ionic strength of the solution is one of the effective factors on cloud point extraction. In this work, the effect of KNO\(_3\) salt as electrolyte was investigated on the process. It is observed that the increasing concentration of salt to the 0.1 M is not effective on the process, but more concentration caused a decrease in absorbance of extracted surfactant-rich phase. This effect might be explained by the additional surface charge when the salt concentration is very high.

**Effect of temperature and equilibrium time on CPE**

It was desirable to employ the shortest equilibration time and the lowest possible equilibration temperature as a compromise between completion of extraction and separation of phases. The dependence of extraction efficiency upon equilibration temperature and time above the cloud point in the range of 30–70 °C and 5–30 min were thoroughly optimized, respectively. It was found that holding the sample solutions for 20 min at 65 °C was quite satisfactory to achieve small volume of the surfactant-rich phase, quantitative extraction and experimental convenience.

**Effect of centrifuge time and rate**

It is necessary to preconcentrate trace amounts of Mg\(^{2+}\) and Ag\(^{+}\) ions with high efficiency in a short period of time. Therefore, CPE on a set of experiments of 15 mL sample at optimum condition by heating 65 °C and centrifuging in various rate and time further cooling in 5 min has been carried out. It was found that centrifugation at 4000 rpm for 15 min separates two phases completely. The enrichment phase did not separate completely, in lower rate and less time centrifugation.

**Calibration, precision and detection limits**

Calibration graph was obtained by preconcentrating 15 mL of several solutions under optimum condition, containing various concentrations of analyte. The process was completed according to the general procedure section, and the absorbance of final solution was measured by FAAS. Linear relationship between the measured absorbance and concentration of Mg\(^{2+}\) or Ag\(^{+}\) ions in the range 0.007–0.2 μg mL\(^{-1}\) was obtained. Table 1 gives the calibration parameters for the proposed CPE method including the linear ranges, the relative standard deviation obtained for 3 analyte samples subjected to the complete procedure, preconcentration and enrichment factors and the limit of detection. It is noted that a calibration curve was obtained without preconcentration in order to calculate the enrichment factor. The enrichment factor was calculated as the ratio of the slopes of calibration curve with and without preconcentration and the preconcentration factor equal to the ratio of the initially volume without preconcentration to finally volume of dissolved preconcentrated phase.\(^{23,36}\)

**Table 1. Analytical characteristics of the metod.**

<table>
<thead>
<tr>
<th>Analyte Conditions</th>
<th>Concentration range (μg mL(^{-1}))</th>
<th>Slope</th>
<th>Intercept</th>
<th>r(^2)</th>
<th>RSD%(^a)</th>
<th>LOD (ng mL(^{-1}))(^b)</th>
<th>Pf and Ef(^c)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Ag(^{+}) Without preconcentration(^d)</td>
<td>2.8-11</td>
<td>0.09</td>
<td>0.9676</td>
<td>0.999</td>
<td>---</td>
<td>---</td>
<td>Pf = 30&lt;br&gt;Ef = 40.57</td>
</tr>
<tr>
<td>Ag(^{+}) With preconcentration(0.06% Triton X-114)(^e)</td>
<td>0.007 - 0.2</td>
<td>3.894</td>
<td>0.027</td>
<td>0.999</td>
<td>0.97</td>
<td>3.00</td>
<td>Ef = 40.57</td>
</tr>
<tr>
<td>Mg(^{2+}) Without preconcentration(^f)</td>
<td>5 - 20</td>
<td>0.012</td>
<td>0.033</td>
<td>0.999</td>
<td>---</td>
<td>---</td>
<td>Pf = 30&lt;br&gt;Ef = 136.09</td>
</tr>
<tr>
<td>Mg(^{2+}) With preconcentration(0.06% Triton X-114)(^g)</td>
<td>0.007 - 0.2</td>
<td>4.490</td>
<td>0.0491</td>
<td>0.999</td>
<td>1.17</td>
<td>1.47</td>
<td></td>
</tr>
</tbody>
</table>

\(^a\)Values in parentheses are the of silver and magnesium concentrations (μg mL\(^{-1}\)) for which the RSD was obtained.

\(^b\)Limit of detection, calculated as three times the standard deviation of the blank signal.

\(^c\)Standard solutions of silver and magnesium in THF medium.

\(^d\)Dilution of the surfactant-rich phase to 500 μl of THF.

\(^e\)Ef and Pf are the Preconcentration factor and Enrichment factors respectively.
Interference study

In the view of high selectivity provided by FAAS, only the interferences studied were those related to preconcentration step, i.e., those cations that may react with ligand and anions that may form complexes with Mg\(^{2+}\) or Ag\(^{+}\) and decrease extraction efficiency. The study of possible interferences caused by cationic species was conducted with some ions under the experimental conditions (which was fixed concentration of 0.133 μg mL\(^{-1}\) of Mg\(^{2+}\) and Ag\(^{+}\), respectively). The results are shown in Table 2. There was no significant interference at a 1:1000 ratio of Mg\(^{2+}\) or Ag\(^{+}\) with the cations Li\(^+\), Na\(^+\), Ag\(^{+}\), Ni\(^{2+}\), Pb\(^{2+}\), Co\(^{3+}\), Ba\(^{2+}\), Ca\(^{2+}\), Zn\(^{2+}\), Cr\(^{3+}\) (for Mg\(^{2+}\)) and Li\(^+\), K\(^+\), Na\(^+\), Ni\(^{2+}\), Mg\(^{2+}\), Ca\(^{2+}\), Pb\(^{2+}\), Zn\(^{2+}\) (for Ag\(^{+}\)) and also the same condition at a ratio of 1:500 Mg\(^{2+}\) or Ag\(^{+}\) with the ions Cu\(^{2+}\), Mn\(^{2+}\), Cd\(^{2+}\), Fe\(^{3+}\), Al\(^{3+}\) (for Mg\(^{2+}\)) and Cu\(^{2+}\), Mn\(^{2+}\), Cd\(^{2+}\), Al\(^{3+}\), Co\(^{3+}\), Hg\(^{2+}\), CH\(_3\)COO\(^{-}\) (for Ag\(^{+}\)).

**Table 2.** Effect of the interference ions on the recovery of the examined metal ions.

<table>
<thead>
<tr>
<th>Interference/analyte ratio</th>
<th>Ion</th>
<th>Analyte</th>
</tr>
</thead>
<tbody>
<tr>
<td>1000</td>
<td>Li(^+), K(^+), Na(^+), Ni(^{2+}), Mg(^{2+}), Ca(^{2+}), Pb(^{2+}), Zn(^{2+})</td>
<td>Ag(^{+})</td>
</tr>
<tr>
<td>500</td>
<td>Cu(^{2+}), Mn(^{2+}), Cd(^{2+}), Al(^{3+}), Co(^{3+}), Hg(^{2+}), CH(_3)COO(^{-})</td>
<td>Mg(^{2+})</td>
</tr>
</tbody>
</table>

**Table 3.** Characteristics of the standard addition curves of Mg\(^{2+}\) and Ag\(^{+}\) ions and recovery of them in water sample.

<table>
<thead>
<tr>
<th>Analyte</th>
<th>Samples</th>
<th>Slope</th>
<th>Intercept</th>
<th>(r^2)</th>
<th>Added (μg mL(^{-1}))</th>
<th>Found (μg mL(^{-1}))</th>
<th>Recovery%</th>
</tr>
</thead>
<tbody>
<tr>
<td>Mg(^{2+})</td>
<td>Well water</td>
<td>4.086</td>
<td>0.011</td>
<td>0.998</td>
<td>---</td>
<td>1.00</td>
<td>0.99</td>
</tr>
<tr>
<td></td>
<td>Spring water</td>
<td>4.035</td>
<td>0.011</td>
<td>0.998</td>
<td>---</td>
<td>1.00</td>
<td>0.98</td>
</tr>
<tr>
<td></td>
<td>Mineral drinking water</td>
<td>4.080</td>
<td>0.010</td>
<td>0.999</td>
<td>---</td>
<td>1.00</td>
<td>0.99</td>
</tr>
<tr>
<td></td>
<td>Tap water</td>
<td>4.491</td>
<td>0.678</td>
<td>0.999</td>
<td>---</td>
<td>1.083</td>
<td>---</td>
</tr>
<tr>
<td>Ag(^{+})</td>
<td>Well water</td>
<td>4.491</td>
<td>0.678</td>
<td>0.999</td>
<td>---</td>
<td>1.00</td>
<td>1.20</td>
</tr>
<tr>
<td></td>
<td>Spring water</td>
<td>4.491</td>
<td>0.689</td>
<td>0.999</td>
<td>---</td>
<td>1.103</td>
<td>---</td>
</tr>
<tr>
<td></td>
<td>Mineral drinking water</td>
<td>4.494</td>
<td>0.701</td>
<td>0.999</td>
<td>---</td>
<td>2.074</td>
<td>2.143</td>
</tr>
<tr>
<td></td>
<td>Tap water</td>
<td>4.482</td>
<td>0.662</td>
<td>0.999</td>
<td>---</td>
<td>1.06</td>
<td>---</td>
</tr>
</tbody>
</table>

Real samples – Evaluation of method

The proposed method was applied for the determination of Mg\(^{2+}\) and Ag\(^{+}\) ions by standard addition technique in a blood serum, urine, radiology film and four different water samples including well water, spring water, tap water and mineral drinking water (Table 3). The results of blood serum and urine samples, as mentioned in the experimental section, are summarized in Table 4. The results of the determination of Ag\(^{+}\) in radiology film sample, as mentioned in the experimental section are listed in Table 5. The recovery of spiked samples is satisfactory and reasonable, which, in view of the high selectivity provided by flame atomic absorption spectrometry, is indicative of the capability of the system in the determination of analytes in real samples. Variety in real samples including four types water, biological and radiological samples express the proposed method is applicable in complex and variously matrices.

**CONCLUSION**

The proposed cloud point extraction method, using ligand as a stable and fairly selective complexing agent offers a simple, rapid, inexpensive and environmentally benign methodology for preconcentration and separation of Mg\(^{2+}\) and Ag\(^{+}\) in aqueous solutions. This method gives very low LOD, good RSD and was applied to the determination of trace amounts of Mg\(^{2+}\) and Ag\(^{+}\) ions in various real samples. In a full comparison of presented results in this
paper with those previously reported, it is found that this method is superior in terms of linear range, detection limits and selectivity. For comparison, the analytical performance data of similar method reported in literature have been listed in Table 5. As it can be seen, the figures of merit of the developed method are comparable or better than the reported methods.

Table 4 Characteristics of the standard addition curves of Mg\(^{2+}\) ion and its recovery in blood serum and urine samples.

<table>
<thead>
<tr>
<th>Samples</th>
<th>Sex</th>
<th>Slope</th>
<th>Intercept</th>
<th>(r^2)</th>
<th>Added (μg mL(^{-1}))</th>
<th>Found (μg mL(^{-1}))</th>
<th>Recovery%</th>
</tr>
</thead>
<tbody>
<tr>
<td>Blood serum</td>
<td>Female</td>
<td>4.458</td>
<td>0.476</td>
<td>0.999</td>
<td>---</td>
<td>2.31</td>
<td>---</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td>---</td>
<td>3.11</td>
<td>100.0</td>
</tr>
<tr>
<td>Urine</td>
<td>Female</td>
<td>4.491</td>
<td>0.784</td>
<td>0.999</td>
<td>---</td>
<td>1.91</td>
<td>---</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td>---</td>
<td>2.91</td>
<td>100.0</td>
</tr>
</tbody>
</table>

Table 5 Characteristics of the standard addition curve of Ag\(^{+}\) ion and its recovery in radiology film sample.

<table>
<thead>
<tr>
<th>Slope</th>
<th>Intercept</th>
<th>(r^2)</th>
<th>Added (μg mL(^{-1}))</th>
<th>Founded (μg mL(^{-1}))</th>
<th>Recovery%</th>
</tr>
</thead>
<tbody>
<tr>
<td>3.867</td>
<td>0.743</td>
<td>0.999</td>
<td>13.30</td>
<td>41.99</td>
<td>99.20</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td></td>
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<td></td>
</tr>
</tbody>
</table>

Table 6. Comparing of the present work with various CPE reported method for determination of Mg\(^{2+}\) and Ag\(^{+}\) ions.

<table>
<thead>
<tr>
<th>Analyte</th>
<th>Method</th>
<th>Ligand</th>
<th>Surfactant</th>
<th>Linear range</th>
<th>D.L</th>
<th>Enrichment factor(Ef)</th>
<th>Ref.</th>
</tr>
</thead>
<tbody>
<tr>
<td>Mg(^{2+})</td>
<td>CPE-FAAS</td>
<td>trizma-chloranilate</td>
<td>Triton X-114</td>
<td>5–220 μg L(^{-1})</td>
<td>0.75 μg L(^{-1})</td>
<td>This work</td>
<td></td>
</tr>
<tr>
<td></td>
<td>CPE-FAAS</td>
<td>4 methyl-3-((1-H-Indol-3-iyl) (phenyl) methyl)-1-H-Indol (MPBIM)</td>
<td>Triton X-114</td>
<td>0.007–0.200 μg mL(^{-1})</td>
<td>1.47 ng mL(^{-1})</td>
<td>136.09</td>
<td></td>
</tr>
<tr>
<td>Ag(^{+})</td>
<td>CPE-FAAS</td>
<td>Bis((1H-benzo[d]imidazol-2yl)methyl)sulfane (BHIS)</td>
<td>Triton X-114</td>
<td>2–400 ng L(^{-1})</td>
<td>1.9 ng mL(^{-1})</td>
<td>39</td>
<td></td>
</tr>
<tr>
<td></td>
<td>CPE-FAAS</td>
<td>2-((2-((1H-benzo[d]imidazole-2-yl)methoxy)phenoxy)methyl)-1H-benzo[d]imidazol (BIMPI)</td>
<td>Triton X-114</td>
<td>28.0–430.0 μg L(^{-1})</td>
<td>10.0 μg L(^{-1})</td>
<td>35</td>
<td></td>
</tr>
<tr>
<td></td>
<td>CPE-FAAS</td>
<td>bis((1H-benzo[d]imidazol-2yl)ethyl) sulfane (BIES)</td>
<td>Triton X-114</td>
<td>1.4 ng mL(^{-1})</td>
<td></td>
<td>42</td>
<td></td>
</tr>
<tr>
<td></td>
<td>CPE-ETAAS</td>
<td>PONPE 7.5</td>
<td></td>
<td>5–100–ng L(^{-1})</td>
<td>1.2 ng L(^{-1})</td>
<td>60</td>
<td></td>
</tr>
<tr>
<td></td>
<td>CPE-FAAS</td>
<td>4 methyl-3-((1-H-Indol-3-iyl) (phenyl) methyl)-1-H-Indol (MPBIM)</td>
<td>Triton X-114</td>
<td>0.007–0.200 μg mL(^{-1})</td>
<td>3 ng mL(^{-1})</td>
<td>40.56</td>
<td></td>
</tr>
</tbody>
</table>

REFERENCES