PREPARATION AND CHARACTERIZATION OF BONDED-PHASES OF CALIXARENESULFONYL-CARBOXYAMIDES IN PARTIAL-CONE CONFORMATION FOR DETERMINATION OF SALBUTAMOL IN LIVESTOCK BY NANO-MEDIATED BONDED-PHASES: NANO-BASKETS OF CALIXARENES IN PARTIAL-CONE CONFORMATION

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ABSTRACT

A new sensitive method for determination of salbutamol, as a nutrient repartitioning agent, in livestock is presented to protect the consumers. Eight nanobaskets of p-tert-calix[4]arene bearing di-[N-(X)sulfonyl carbamamide] and di-(1-propano) in partial-cone conformation were synthesized and were used to prepare bonded phases of HPLC–UV. The new synthesized bonded-phases were characterized and optimized, the bonding interactions of solute and stationary phases were examined and the main interactions were reported. The results revealed that for the best bonded-phase, the LOD and LOQ were 0.02 and 0.06 μg/mL, respectively.

Keywords: Bonded-Phase; Partial-Cone Conformation; Calixarene; Salbutamol; nanobasket.

1. INTRODUCTION

Salbutamol (albuterol) is the international nonproprietary (United States) adopted name for 4-[(2-(tert-butylamino)-1-hydroxyethyl)-2-(hydroxymethyl)phenol (Figure 1), which is used for the treatment of asthma in both humans and animals, is a synthetic β₂-agonist. In high doses, salbutamol exhibits a metabolic effect, which results in an increase in muscle mass and a decrease in adipose tissue. So, the compound is also misused as nutrient reparationing agent in livestock by diverting nutrients from fat deposition in animals to the production of muscle tissues [1]. This misuse had caused some severe accidental poisonings in humans [2,3]. Therefore, all β₂-agonists are banned for growth promotion in animal production. In order to protect consumers, specific and sensitive methods for the identification and quantitation of salbutamol in meat and other food are required.

Figure 1. The chemical structure of salbutamol.

Nano-baskets of calixarenes and calixcrowns are a versatile class of macrocycles, which have been subject to extensive research in development of many extractants, (using gas chromatograph, Teif Gostar Faraz Co.) transporters and stationary phases over the past four decades [4-6]. Functionalization of calix[4]arenes at both the upper rim and the lower rim has been extensively studied [7-9]. Attaching donor atoms to the lower rim of a calix[4]arene can improve the binding strength of the parent calixarene dramatically [10-12]. Calixarene-bonded stationary phases are preferable to use as mobile–phase additives, because the UV detection of analytes is prevented by strong absorbance of calixarenes [13-15]. Additionally, poor solubility of most calixarenes precludes their applications as additives in aqueous eluents. With the advent of various functionalization methods of calixarenes, more applications have been reported for calixarene bonded-phases. The resulting interactions of different calixarenes stationary phase influence the retention factors and improve the selectivity of the solutes. The modification of the calixarenes, for instance, by varying the conformations, ring size, and substituents, enable a more enhanced interaction spectrum and can improve the specificity for guest molecules [16-21]. Table 1 summarizes the recent research carried out using calixarenes in liquid chromatography other applications of calixarenes are reported in microextraction [44], desalination [45] and drug developments [46].

In the present work, the synthesis of eight derivatives of p-tert-calix[4] arene bearing di-[N-(X)sulfonyl carbamamide] and di-(1-propano) in partial-cone conformation and the preparation of its bonded silica stationary phase were described (X= Phenyl (01,05), p-CH₂Phenyl (02,06), p-OHPhenyl (03,07), p-NO₂Phenyl (04,08)). A new method for determination of salbutamol in the livestock meat samples was set up by using the new calix[4]arene-based column. The chromatographic performance using salbutamol as well as the influence of methanol content (in mobile phase) on the chromatographic behavior of the solutes was investigated. Figure 2 shows the expanded chemical structure of derivatives 01-08.

Figure 2. Chemical structure of derivatives 01-08.

2. Experimental Procedure

2.1. Materials and Apparatus

All solvents and analytes used were of analytical grade and obtained from Merck Chemical Company unless specially mentioned. Silica gel (with particle size of 5 μm, pore size of 100 Å and specific surface area of 300 m²/g), HPLC-grade methanol (MeOH) and γ-glycidoxypropyltrimethoxysilane (KH–560) were used. A phosphate buffer (0.5%, w/w, pH 4.5) was prepared by mixing KH₂PO₄ with ultra–high quality pure water, and filtered through a 0.45 μm filter before use. Water was purified by using Milli–Q purification equipment.

The stock solution of Salbutamol (250 mg/mL) was prepared by dissolving the reference substance in MeOH, and stored in the refrigerator. The standard working solutions were prepared by diluting aliquots of the stock solution to obtain concentrations ranging from 0.1 to 2.0 μg/mL. The calibration graph was constructed by plotting the peak areas obtained at wavelength 243 nm versus the corresponding injected concentrations. The samples were treated according to the following method: tissue samples (2.000 g) was transferred to a polypropylene centrifuge tube (50 mL) with stopper, ammonium acetate (10
mL, 20 mmol/L) buffer solution was added to the tube, then β-glucuronidase–aryl sulfatase (50 μL, containing 134600 U/mL β-glucuronidase and 5200 U/mL aryl sulfatase) was added; after that, oscillated on the vortex oscillator (2 min) and sonicated (20 min) by using a ultrasonic processor, then put into a incubator (16 h, 37°C). After the tube was centrifuged at 4000r/min (10 min), the supernate was transferred into another tube. Then, chloroform (5.0 ml) was added, oscillated and sonicated again. The supernate was applied to Waters Oasis MCX cartridge, which was activated with methanol (5mL), water (5mL), and followed by hydrochloric acid (5 mL, 0.01 mol/L) solution. The cartridge was washed with ethyl acetate/ammonium hydroxide (6 mL, 97:3, v/v), the eluent was collected into a glass tube, and evaporated until dryness under a stream of nitrogen at 40°C water bath, the residue was reconstituted in 0.2% formic acid in water/acetonitrile (1.0 mL, 90:10, v/v). The resulting solution was filtered through 0.22 μm filter and the filtrate (10 μL) was ready for analysis.

Table 1. The list of calixarenes used as stationary phases.

<table>
<thead>
<tr>
<th>Type of calixarene</th>
<th>Analytes were used</th>
<th>References</th>
</tr>
</thead>
<tbody>
<tr>
<td>calix[4,6,8]arene</td>
<td>PAHs</td>
<td>[22]</td>
</tr>
<tr>
<td>calixBzCl, calixBzF, calixBzNO2, calixBn, calixBph, calixBnOMe</td>
<td>pentylbenzene, butylbenzene, triphenylene, o-terphenyl, caffeine, phenol, benzyalmine</td>
<td>[23]</td>
</tr>
<tr>
<td>C-tetraundecylcalix[4]resorcinarene</td>
<td>cytosine, uracil and thymine</td>
<td>[27]</td>
</tr>
<tr>
<td>calix[8]arene</td>
<td>tricyclic neuroleptic</td>
<td>[28]</td>
</tr>
<tr>
<td>calix[4]arrenes</td>
<td>benzene or uracil derivatives</td>
<td>[29]</td>
</tr>
<tr>
<td>caltrex BIE</td>
<td>paracetamol, caffeine and acetyl salicylic acid</td>
<td>[30]</td>
</tr>
<tr>
<td>caltrex AIII</td>
<td>celecoxib</td>
<td>[31]</td>
</tr>
<tr>
<td>p-tert-butyl-calix[8]arene</td>
<td>aromatic compounds</td>
<td>[33]</td>
</tr>
<tr>
<td>caltrex AI, caltrex AII, caltrex AIII, caltrex BI, caltrex BII and caltrex BIII</td>
<td>PAHs, phenols, substituted aromatics, xanthines, barbituric acid, benzoic acid esters</td>
<td>[35]</td>
</tr>
<tr>
<td>calix[4]arene tetradiethylamide</td>
<td>amino acid ester hydrochlorides</td>
<td>[36,37]</td>
</tr>
<tr>
<td>calix[4]resorcinarene and calix[4,6,8]arene</td>
<td>flupentixol, chlorprothixene, clopenthixol, and doxepin</td>
<td>[38]</td>
</tr>
<tr>
<td>p-tert-butylcalix[4,6,8]arene</td>
<td>disubstituted aromatics, uracil derivatives, and estradiol epimers</td>
<td>[39]</td>
</tr>
<tr>
<td>p-tert-butylcalix[4,6,8]arene</td>
<td>peptide bond isomers of proline</td>
<td>[40]</td>
</tr>
<tr>
<td>p-tert-butyl-calix[4]arene</td>
<td>nucleosides, PAHs and bases</td>
<td>[41]</td>
</tr>
<tr>
<td>calix[6]arene</td>
<td>nucleosides and PAHs</td>
<td>[42]</td>
</tr>
<tr>
<td>p-tert-butyl-calix[4]arene</td>
<td>basic aromatic compounds, polar and non-polar aromatic compounds, and PAHs</td>
<td>[43]</td>
</tr>
</tbody>
</table>

Chromatographic analyses were carried out by using an Agilent 1200 series system equipped with a 1200 model quaternary pump, a G1314A model Multiple Wavelength UV–vis detector, a G1316A model thermostated column compartment, a 1322A model vacuum degasser, and an Agilent Chemstation B.03.02 Patch data processor. The homemade calixarene column was filled using a packing machine under the pressure of 50 MPa. An Eclipse XDB–C18 column (Agilent, 150 mm × 4.6 mm i.d., 5 μm) was used as a comparison with the homemade calixarene column.

2.2. Preparation of Stationary Phases

Figure 3 shows the preparation process of a new calix[4]arene–bonded silica gel stationary phase 01. Active silica gel (5.0 g) was suspended in dry toluene (50 mL, freshly distilled), and then KH–560 (6.0 mL) and triethylamine (1.0 mL, used as a catalyst) was added to this suspension. The mixture was stirred and heated to 80°C under the protection of nitrogen atmosphere (8 h). After the reaction finished, the solid was filtered by 1.5 μm filter, and washed in sequence with toluene and acetone, then dried at 80°C under vacuum (8 h). Finally, γ-glycidoxypropyl bonded silica gel (γ-GBSG) was obtained and used as a precursor in the following reaction.
Figure 3. Preparation of a calix[4]arene–bonded silica gel stationary phase 01.

Each of the para-tert-butyl calix[4]arenes 01-08 (3.0 g, 3.1 mmol), NaH (0.6 g), and toluene (60 mL, freshly distilled) were stirred at 80°C for 30 min, then the supernatant liquid was transferred to a 100 mL three-neck flask, γ-GBSG (3.0 g) was added, and the mixture was refluxed with the catalyst for 48 h. The whole process was carried out under nitrogen atmosphere. After the reaction finished, the product was filtered and washed in sequence with toluene, acetone, methanol and distilled water. Subsequently, stationary phases were obtained and dried at 100°C under vacuum for 8 h, then cooled to room temperature in a desiccator.

2.3. Characterization of Stationary Phases

The characterization of the developed stationary phases was carried out by elemental analysis, IR and thermal gravimetric analysis. Thermal gravimetric analysis (TGA) was carried out with a Shimadzu DT–60 thermal analyzer, the analysis was performed from 40°C to 650°C at heating rate of 10°C/min in argon atmosphere with a gas flow rate of 20 mL/min.

Table 2 shows the elemental analysis results of γ-GBSG and stationary phases. The result indicates stationary phases own higher content of carbon, nitrogen and sulfur than that of γ-GBSG, which confirmed that the calixarene was successfully immobilized onto the silica gel. The bonded amount of calix[4]arene derivatives onto the silica gel was calculated by subtracting that of γ-GBSG. According to the carbon content of the bonded silica gel stationary phases, the resulting stationary phases contain about 10% carbon corresponding 0.05 mmol calix[4]arene per gram of silica-gel.

Moreover, the bonded stationary phases were characterized by infrared spectroscopy. From IR spectra, the characteristic absorption band of the benzene ring appears around 1625, 1552 and 1486 cm⁻¹, peaks around 2960 and 2865 cm⁻¹ are assigned to C–H stretching frequency. The peaks around 1712 cm⁻¹ are assigned to the absorption frequency of the –C=O and –C–N groups. The peaks around 1105 cm⁻¹ correspond to the groups of Si–O–Si and C–O–C. All IR spectra indicate that the organic ligands were bonded onto silica gel.

The thermal stability of γ-GBSG and stationary phases has been investigated by TG analysis. The results show that the temperatures of weight loss for γ-GBSG and stationary phases are both more than 300°C. It indicates that the new bonded phases possess high thermal and chemical stability. Moreover, the weight losses in 25–650°C were 7 and 13 % for γ-GBSG and stationary phases, respectively, which was in line with the results of elemental analysis.

2.4. Chromatographic Procedures

Eight columns (150 mm × 4.6 mm i.d.) were packed with modified calix[4]arene–silica gels according to a slurry packing procedure by using methanol as the displacing agent (50 MPa, 1 h). The mobile phases used were methanol–water and methanol–phosphate buffers (0.5%, w/w, pH 4.5). Analytes were dissolved in the mobile phase at the concentration in range of 5–100 μg/mL, and solution (20 μL) was injected into the chromatographic column. The void time (t_v) for the calculation of the retention factor was determined by injecting sodium nitrate (0.05 M, NaNO₃) at UV detection 210 nm, with MeOH–H₂O (70/30, v/v) as mobile phase. All measurements were carried out at 25°C and repeated three times.

3. RESULTS AND DISCUSSION

In this section, the column factors are determined and followed by description of analytical results and analyte: bonded-phase interactions. Finally, the effect of organic modifier (methanol) is discussed.

3.1. Column Factors

The retention factors (k') and the separation factors (α) of eight prepared columns were calculated and listed in table 3. Separation factors were determined using the closest signal to the salbutamol’s signal. This result indicates that new stationary phases can behave as excellent reversed-phase performance and the hydrophobic interaction is one of the factors playing a role in the separation of analyte.

### Table 3. Column factors for separation of salbutamol on bonded-phases 01-08.

<table>
<thead>
<tr>
<th>Bonded phase</th>
<th>Retention factor</th>
<th>Separation factor</th>
</tr>
</thead>
<tbody>
<tr>
<td>01</td>
<td>2.2</td>
<td>1.7</td>
</tr>
<tr>
<td>02</td>
<td>1.9</td>
<td>1.9</td>
</tr>
<tr>
<td>03</td>
<td>3.1</td>
<td>1.7</td>
</tr>
<tr>
<td>04</td>
<td>3.1</td>
<td>1.8</td>
</tr>
<tr>
<td>05</td>
<td>2.1</td>
<td>1.7</td>
</tr>
<tr>
<td>06</td>
<td>1.8</td>
<td>1.7</td>
</tr>
<tr>
<td>07</td>
<td>2.9</td>
<td>1.7</td>
</tr>
<tr>
<td>08</td>
<td>3.0</td>
<td>1.7</td>
</tr>
</tbody>
</table>

3.2. Analytical Results

The graph of the peak area (y) against concentration (x, μg/mL) proved linear in the range from 0.1 to 20.0 μg/mL and the regression coefficient R² = 0.9996. The limit of detection (LOD), was defined as the injected quantity...
Effect of the methanol content of mobile phases on the retention factors playing a role in the separation of salbutamol. However, it is obvious that the new stationary phases can behave as an excellent reversed-phase performance and the hydrophobic interaction is one of the factors responsible for the retention behavior of salbutamol, and the columns exhibited good hydrophobic interaction properties except for columns 02, 06.

In figure 4, the upper chromatograms (st.) depict the standard solution containing salbutamol (0.2 µg/mL) and the other traces show the chromatograms of the following samples: pork (P), pork casing (PC), beef (B), beef casing (BC), mutton (M) and mutton casing (MC), respectively.

3.3. Column: Analyte Interactions

The aromatic rings, X groups (defined previously) and the cavity of the calix[4]arene play the additional roles on the separation mechanism for the salbutamol. These additional interactions are possibly inclusion complexation, π–π, hydrogen bonding, and hydrogen bonding interactions.

In all bonding phases 01-08, the hydrophobic interaction was mainly responsible for the retention behavior of salbutamol, and the columns exhibited good hydrophobic interaction properties except for columns 02 and 06. At the same time, from the chemical structures and the chromatograms it was obvious that salbutamol can easily form hydrogen bond via it's OH and NH functional groups with the relevant groups on the calixarene-bonded phases. Moreover, the electronic-rich cavity of calixarene can attract the functional groups mentioned above on the salbutamol, which seems to be more for bonded phases 02 and 06.

3.4. Effect of Organic Modifier

Figure 5 illustrates the plot of logarithmic retention factor of salbutamol against the volume percentage of methanol in the mobile phase. As methanol content of the mobile phase increasing, the retention factors are decreasing. This result indicates that the new stationary phases can behave as an excellent reversed-phase performance and the hydrophobic interaction is one of the factors playing a role in the separation of salbutamol. However, it is obvious that the relationship between log k' and the content of methanol is not linear. Moreover, by increasing the methanol contents to certain values, the relationship line of log k' and methanol content tends to be flaten, which means that there exist other interactions in the separation process when the methanol content was in the low levels. Hence, it indicates that hydrophobic was not the only factor in the separation of the solute and the X functional groups may also be responsible for the retention behavior. According to figure 5, when the methanol content was in the low levels, all bonded phases show different k' values; while at the higher levels of methanol, they depict just two k' values. These results revealed that inclusion complexation, π–π and hydrogen bonding interactions in bonded phases 04, 08 are the most and followed by the order of 02, 06 < 01, 05 < 03, 07 < 04, 08.

4. CONCLUSIONS

Eight new p-tert-butylcalix[4]arene columns containing sulfonyl carboxamidic groups in partial-cone conformation were prepared and characterized by FTIR, elemental and thermal analysis. The chromatographic behaviors of new developed stationary phases were investigated towards salbutamol using six samples of livestock meat. By increasing the methanol content of the mobile phase, the retention factors were decreased and showed that various chromatographic retention mechanisms occur in the separation of the above analyte, such as π–π interaction, hydrogen bonding interaction and inclusion complexation. The columns were successfully used for the analysis of salbutamol pork, pork casing, beef, beef casing, mutton, and mutton casing except for columns 02 and 06.

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